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Group Nr.:

Date:

# LABORATORY PROTOCOL Nr.2

**Topic: Direct detection and antibiotics**

**Task Nr.1: Cultivation**

Principle:

Cultivation is based on the ability of microorganisms to multiply on inanimate media.

Conclusion:

McConkey agar	
Blood agar	

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### Task Nr.2: Counting of colonies

Principle:

One colony will grow from one microbe cell in case the cells on the agar surface are well separated. This is achieved by appropriate dilution of the sample.

Procedure:

Add 0.1 ml of bacterial suspension to test tube 1 with 9.9 ml of phys. saline solution, after mixing transfer 0.1 ml suspension from test tube 1 to test tube 2, then from test tube 2 to test tube 3. From tubes 2 and 3, inoculate 0.1 ml of suspension on agar, spread it over the whole surface by loop. Incubate for 24 hours at 37°C and check the plate.

### Microbial concentration (K) - calculation

Volume (V), dilution(D), and numbers of colonies (N)

$$K = N/V \times D$$

Conclusion:

Tube	Dilution	Numbers of colonies	Microbial concentration
2			
3			

### Task Nr.3: Determination of susceptibility of bacterial strain using disc diffusion method

Principle:

Determination of susceptibility of non-fastidious bacteria to antibiotics using antibiotic discs.

Procedure:

Incubate for 18 hours at 37°C and read the inhibition zone.

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Antibiotic discs	Cephoxitin 30 µg	Erythromycin 15 µg	Clindamycin 2 µg	Tetracyclin 30 µg	Chloramphenicol 30 µg	Gentamicin 10 µg
Zone size						
Breakpoint	C ≥22 / R <22	C ≥21 / R <18	C ≥22 / R <19	C ≥22 / R <19	C ≥18 / R <18	C ≥18 / R <18
Evaluation						

Conclusion:

#### Task Nr.4: Nitrocefin test

Principle:

We demonstrate the production of beta-lactamase, which cleaves the beta-lactam ring of antibiotics.

Procedure:

Moisten a strip of filter paper with a solution of nitrocefin and inoculate by the test strain. Check the color reaction.

Conclusion: