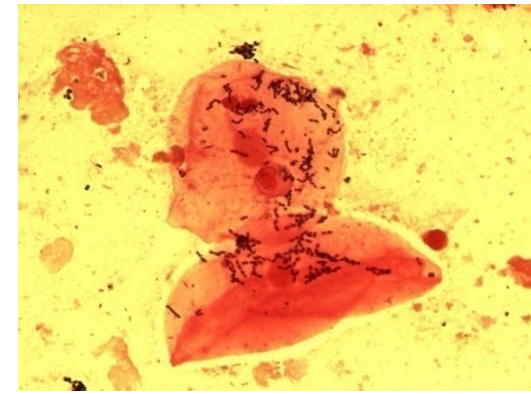
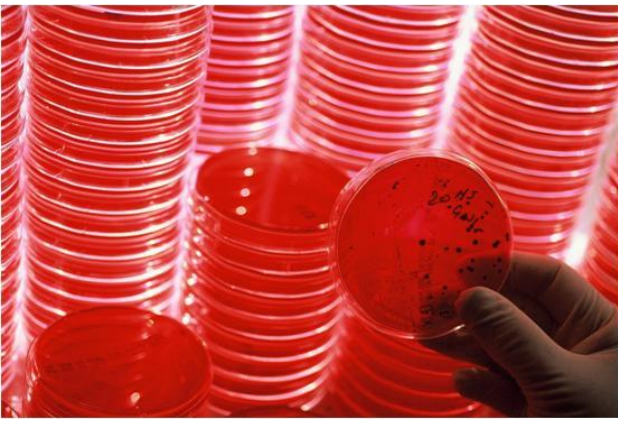


# Molecular microbiology

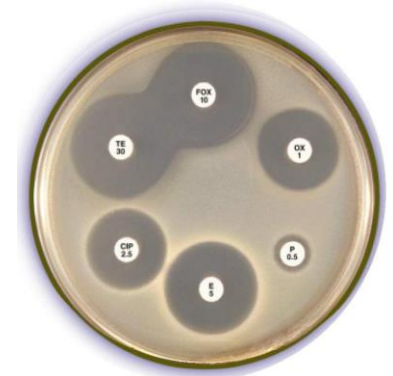


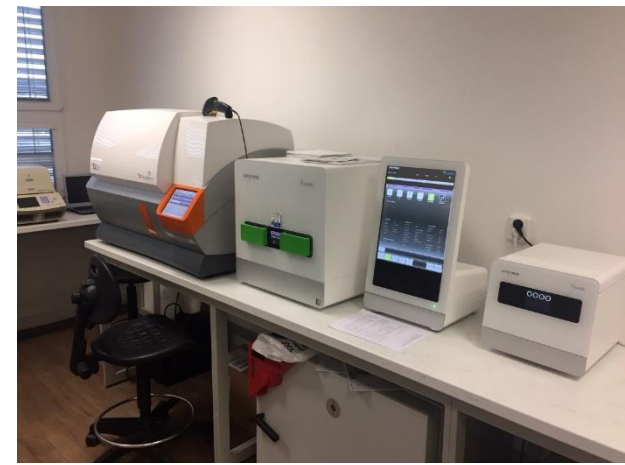
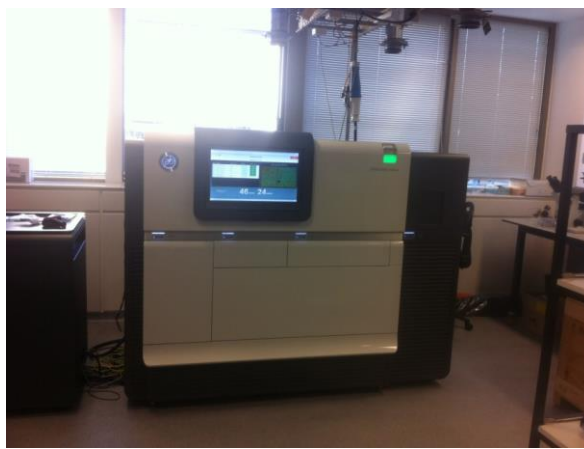
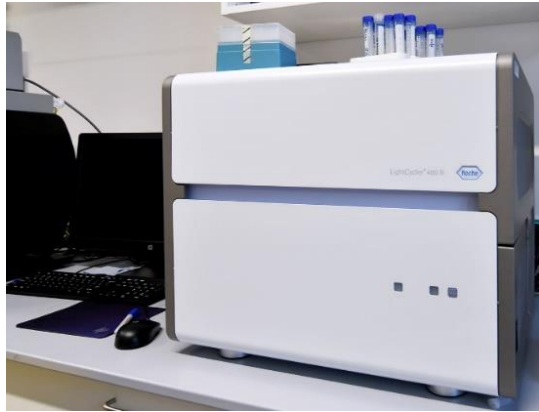
Pavel Drevinek



## Traditional microbiology

- microscopy
- culture, identification and antibiotic susceptibility tests
- antigen detection
- serology and antibodies

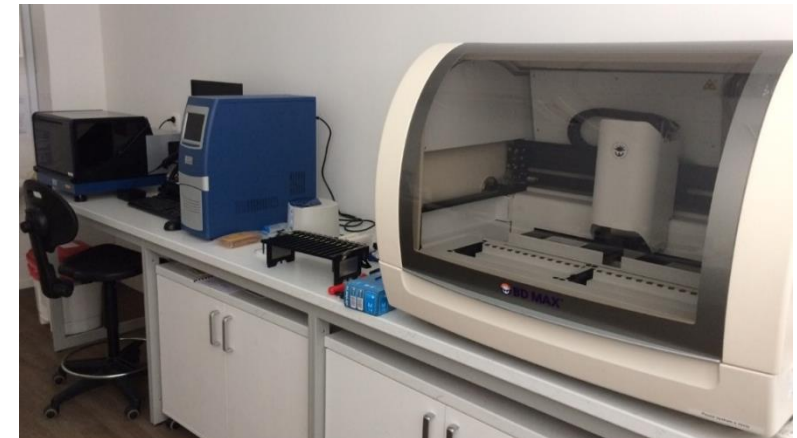




## Molecular microbiology

DNA or RNA analysis for the purpose of:

- diagnostics
- detection of virulence genes
- detection of antimicrobial resistance genes
- epidemiology



# To detect DNA or RNA for diagnostic purposes

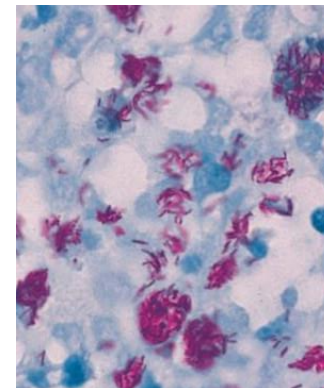
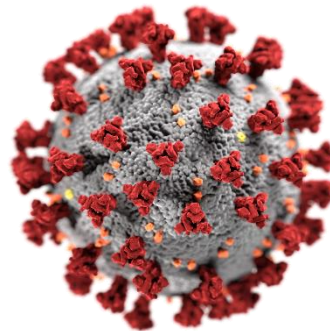
## Advantages

Diagnostics of culture negative infections

- non-culturable agents, slow growing, „fastidious“
- detection while on antibiotic therapy
- high sensitivity

Speed

Quantification (viral load)



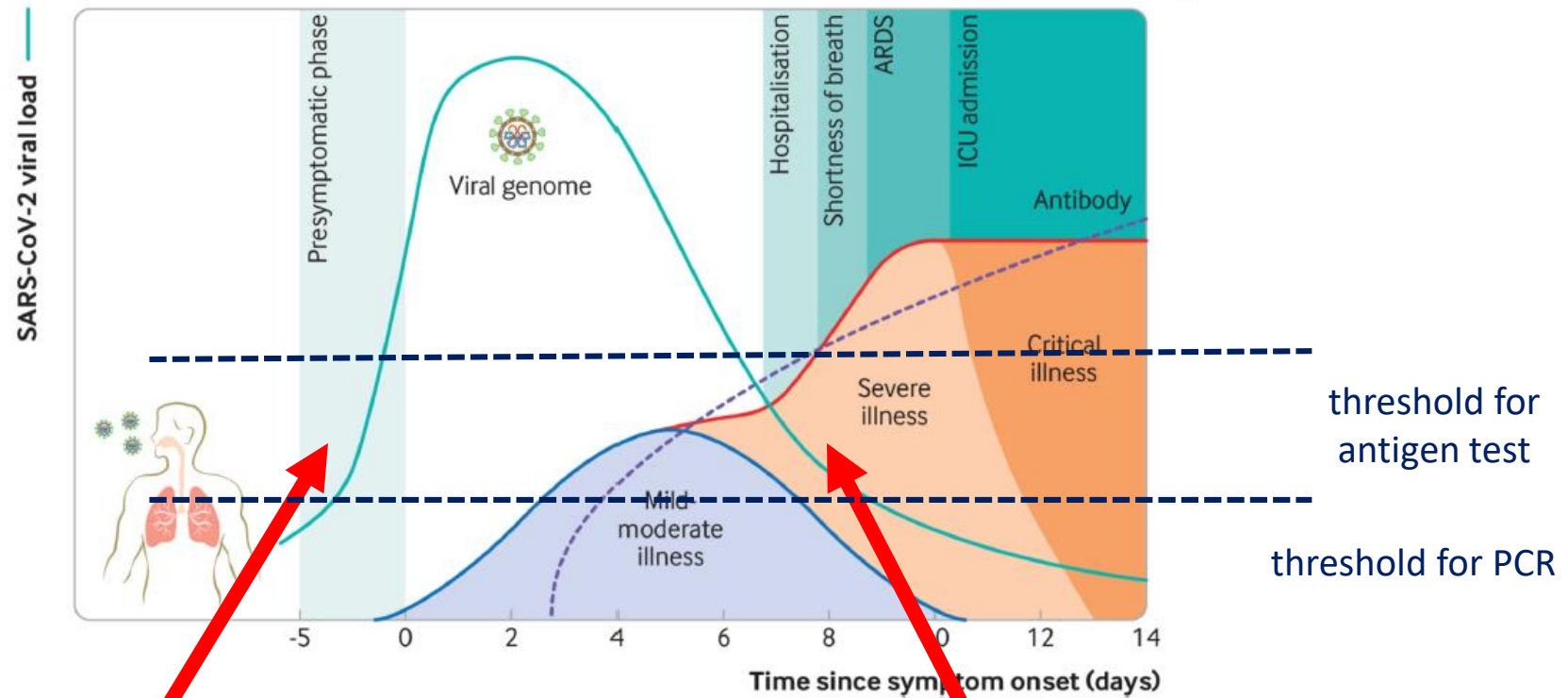
## To detect DNA or RNA for diagnostic purposes

### Drawbacks

Pitfalls in interpretation, invisible at first sight:

- does the detected DNA originate from a viable agent?
- positivity: contaminant, bystander or pathogen?

# PCR and "false positivity"



- early (and desirable) detection (only with PCR)

- persistent, artificial, not clinically relevant positivity



DNA diagnostics



preselected single  
microorganism

pathogen specific PCR

- SARS-CoV-2
- *Bordetella pertussis*
- EBV (quantity)
- PVL
- ...



DNA diagnostics



preselected single microorganism



preselected multiple microorganisms

multiplex, syndromic, panel PCR

**Respiratory infections atypical**

- Mycoplasma pneumoniae*
- Chlamydia pneumoniae*
- Chlamydia psittaci*
- Legionella pneumophila*
- Pneumocystis jirovecii*
- Cryptococcus neoformans*

**Sexually transmitted diseases**

- Neisseria gonorrhoeae*
- Chlamydia trachomatis*
- Mycoplasma genitalium*
- Mycoplasma hominis*
- Ureaplasma urealyticum*
- Ureaplasma parvum*
- Trichomonas vaginalis*

**Gut infections**

- Salmonella*
- Campylobacter*
- Shigella/ E. coli (EIEC)*
- Shiga toxin (EHEC)*
- Clostridium difficile (toxin A,B)*
- Aeromonas*
- Yersinia*
- Cryptosporidium spp.*
- Sapovirus
- Rotavirus
- Norovirus
- Adenovirus
- Astrovirus
- Giardia intestinalis*
- Entamoeba histolytica*



# POCT

point-of-care tests or bedside or near-patient testing

- instant availability of results
- run by personnel not qualified for laboratory work

## Single agents:

- SARS-CoV-2
- RSV
- influenza



by 15 mins

600 Kc

- *M. tuberculosis*
- MRSA
- *C. difficile*



45 to 80 mins

1500 Kc

# POCT

point-of-care tests or bedside or near-patient testing

- instant availability of results
- run by personnel not qualified for laboratory work

Multiple agents:

influenza A  
influenza B  
RSV  
SARS-CoV-2



60 mins

# POCT

point-of-care tests or bedside or near-patient testing

- instant availability of results
- run by personnel not qualified for laboratory work

Multiple agents:

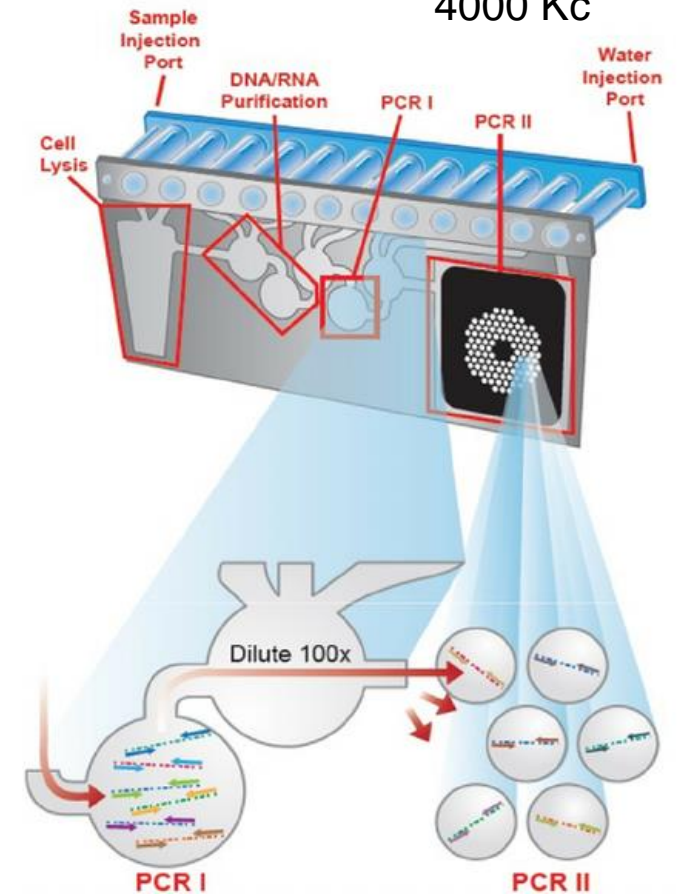
<i>S. pneumoniae</i>	enteroviruses
<i>N. meningitidis</i>	HSV1
<i>H. influenzae</i>	HSV2
	VZV
<i>S. agalactiae</i>	CMV
<i>E. coli</i>	HHV6
<i>L. monocytogenes</i>	parechovirus
	<i>Cryptococcus neoformans</i>



60 mins

## The FilmArray Pouch

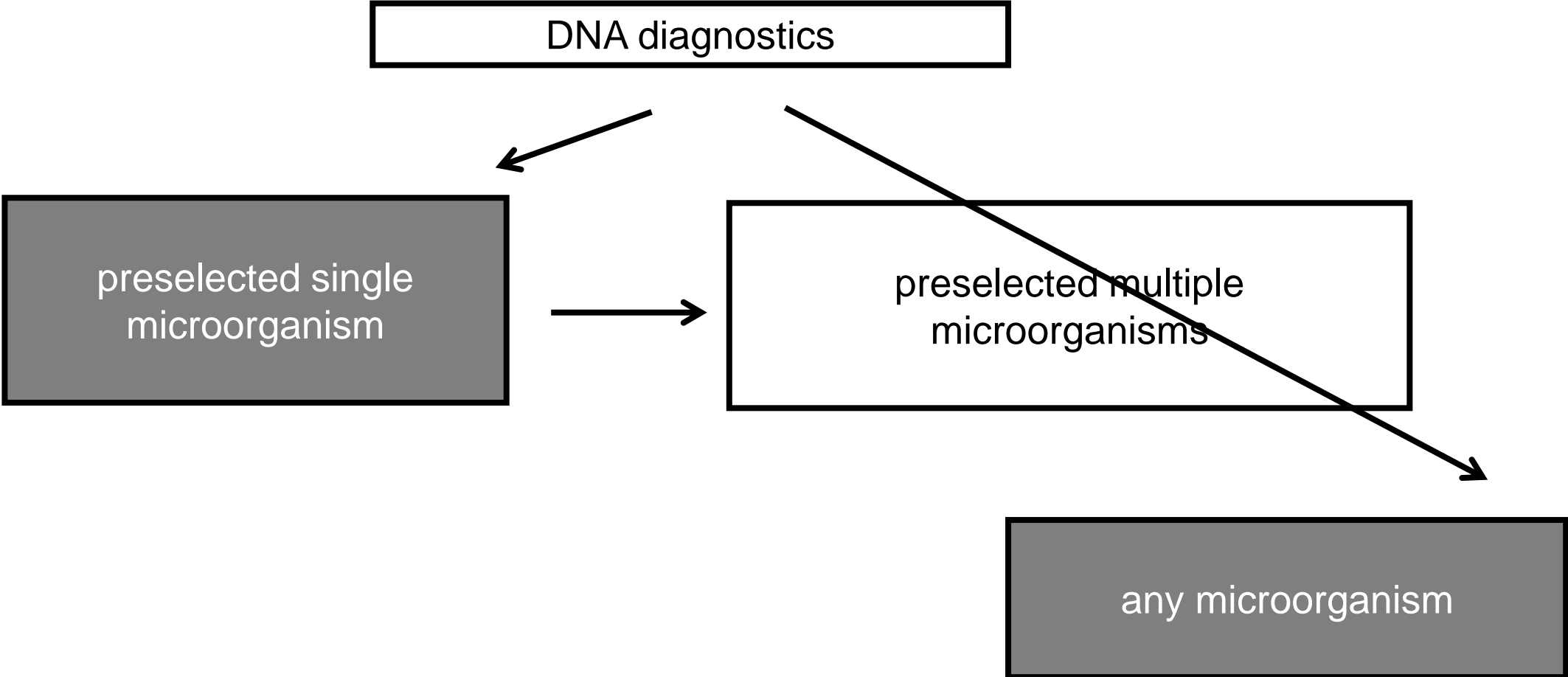
4000 Kc



## Pneumonia panel

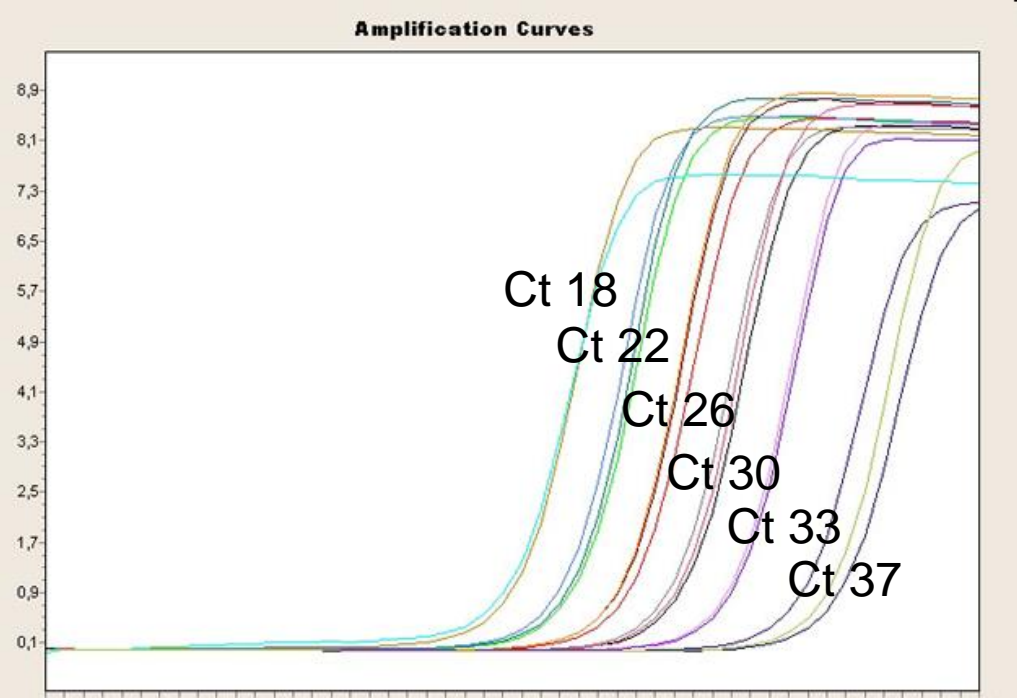
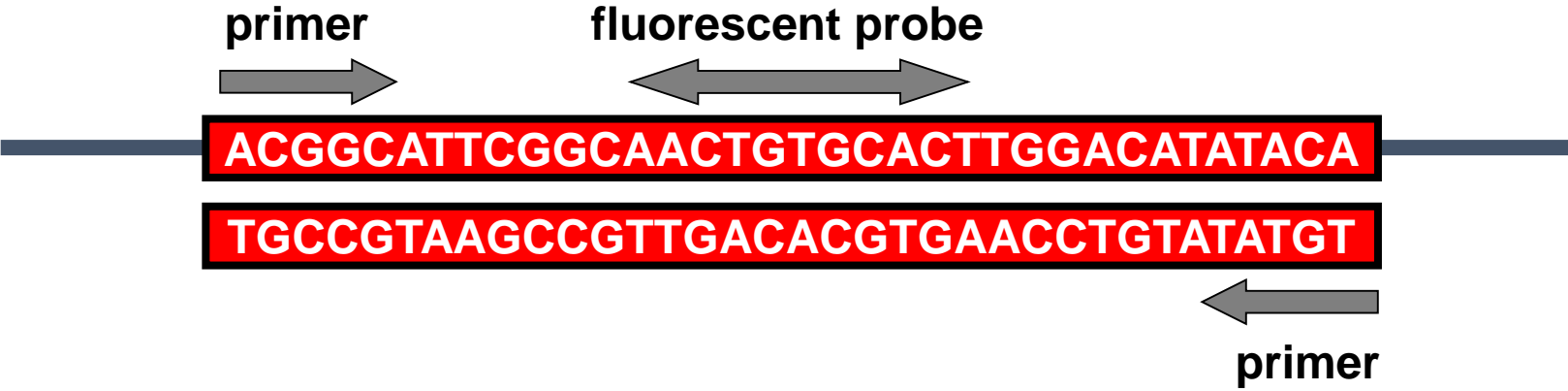
influenza A	<i>Klebsiella pneumoniae</i>
influenza B	<i>Klebsiella oxytoca</i>
RSV	<i>Klebsiella aerogenes</i>
rhinoviruses/enteroviruses	<i>Moraxella catarrhalis</i>
parainfluenza	<i>Serratia marcescens</i>
adenoviruses	<i>Staphylococcus aureus</i>
metapneumovirus	<i>Streptococcus pneumoniae</i>
seasonal coronavirus	<i>Streptococcus pyogenes</i>
SARS-CoV-2	<i>Streptococcus agalactiae</i>
<i>Mycoplasma pneumoniae</i>	
<i>Chlamydia pneumoniae</i>	
<i>Legionella pneumophila</i>	
<i>Acinetobacter baumannii</i>	
<i>Pseudomonas aeruginosa</i>	
<i>Enterobacter cloacae</i>	
<i>Proteus spp.</i>	
<i>Escherichia coli</i>	
<i>Haemophilus influenzae</i>	

CTX-M  
IMP  
KPC  
NDM  
VIM  
OXA-45-like  
mecA/C and MREJ



How to actually „design“ PCR  
to make it working according to our wishes

# Molecular microbiology: targetting nucleotide sequence of a microbe



Ct value = PCR cycle when the signal starts to grow

the higher the Ct value, the less agent is detected in the sample

Does the patient have a whooping cough?

(you need to detect only *Bordetella pertussis*; ignore others)

DNA diagnostics



```
graph TD; A[preselected single microorganism] --> B[DNA diagnostics]
```

preselected single  
microorganism



the target sequence for the primers must be unique to bordetella



Joint infection. What is the cause?

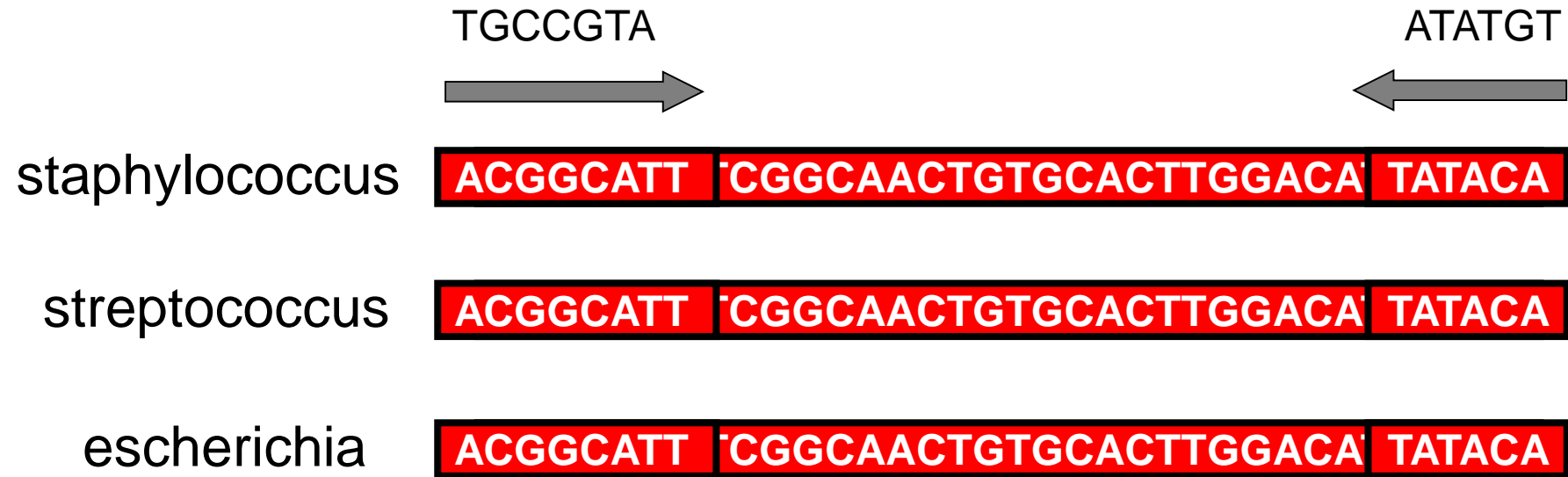
(you need a tool that enables to detect any bacteria)

DNA diagnostics



any microorganism

the target sequence for the primers is present in all bacteria



PCR positivity

+

sequencing



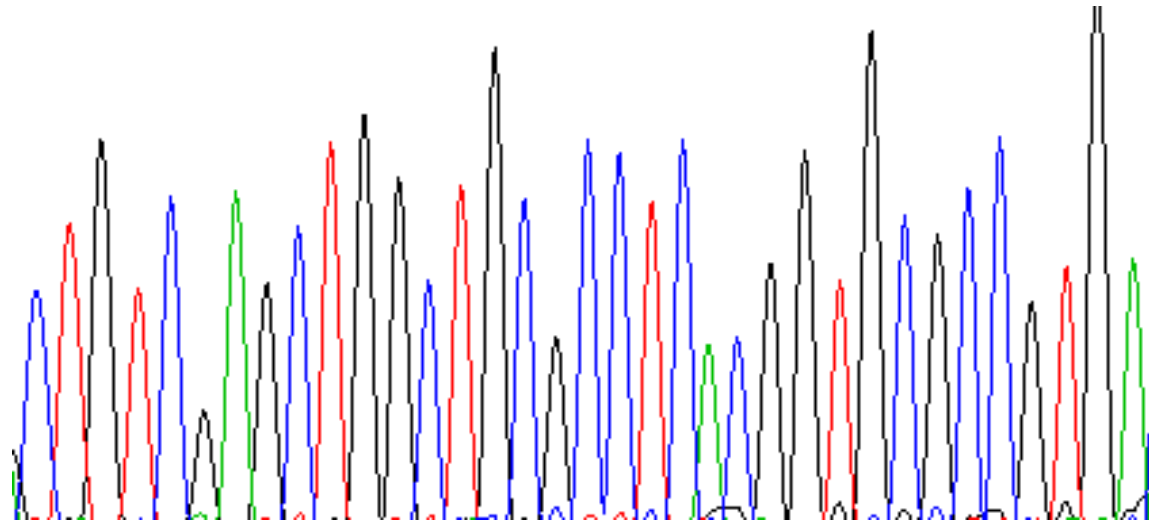
DETECTION OF BACTERIA,  
but which one?



IDENTIFICATION  
on a species level

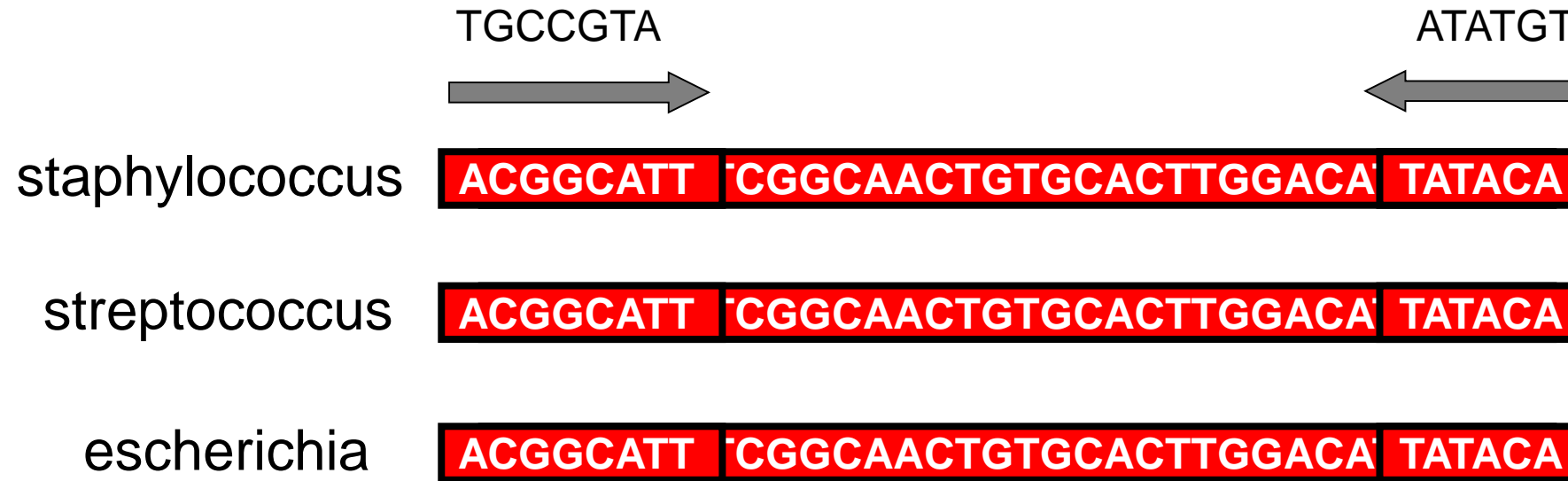
# PCR product sequencing:

| C T G T C G A G C T G G C T G C G C C T C A C G G T G C G C C G T G A C → *S. aureus*

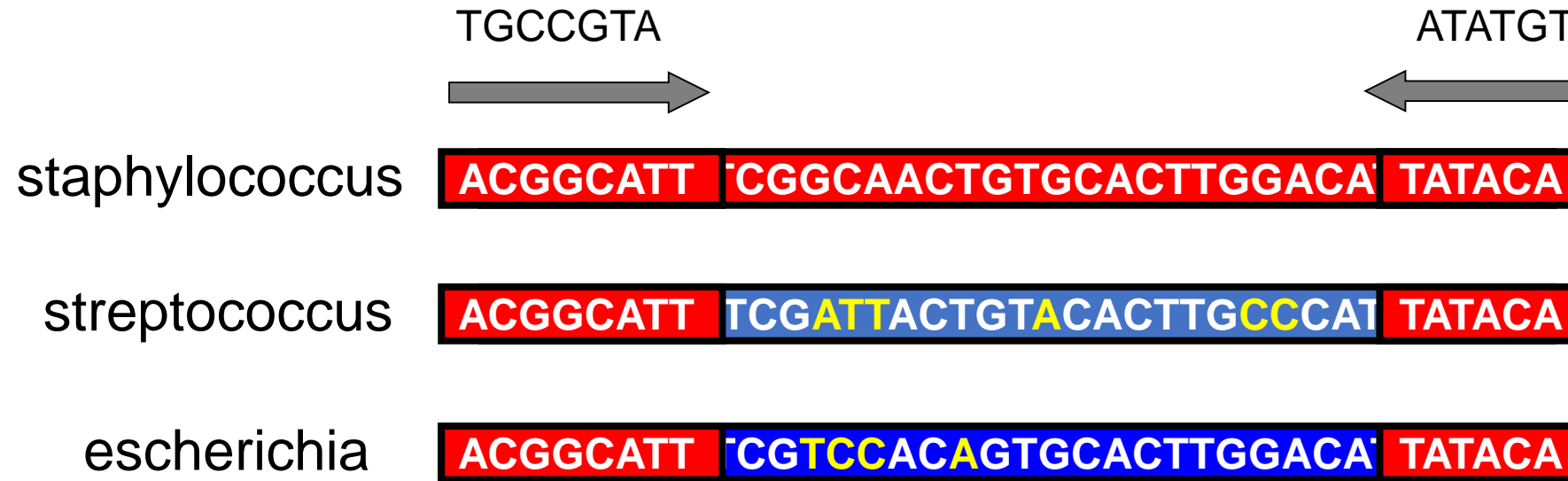


C T G A

the target sequence for the primers is present in all bacteria

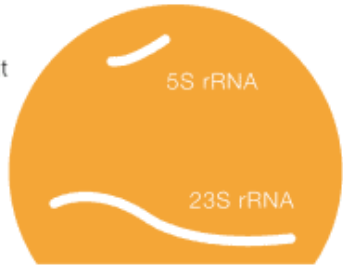


the target sequence for the primers is present in all bacteria



# Prokaryotic Ribosome

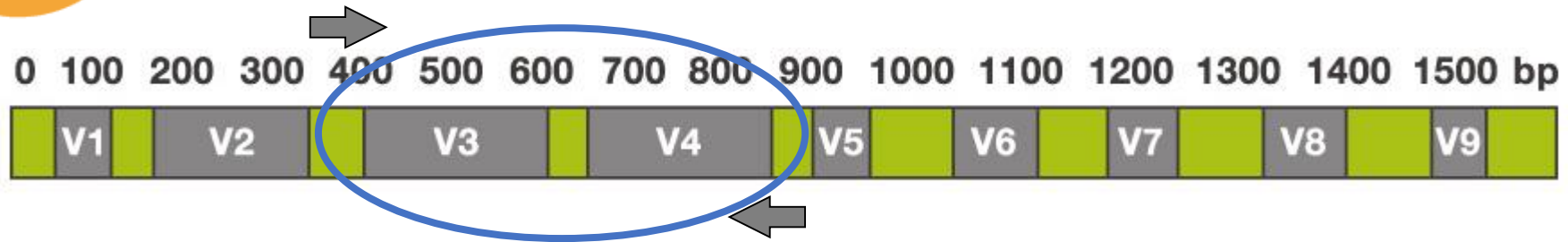
Large Subunit (LSU)



Small Subunit (SSU)



16S rRNA gene



Note:

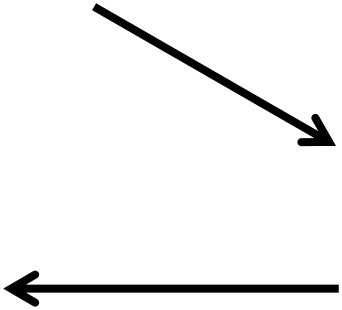
panbacterial PCR to be done from primary sterile material only



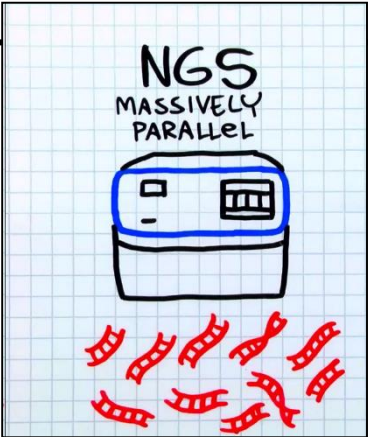
DNA diagnostics

all microorganisms  
(metagenome)

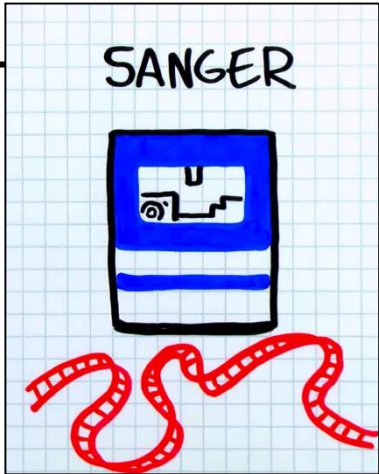
any microorganism



**Method**  
Massive parallel sequencing  
(NGS)



**Method**  
PCR 16S rRNA  
and Sanger sequencing

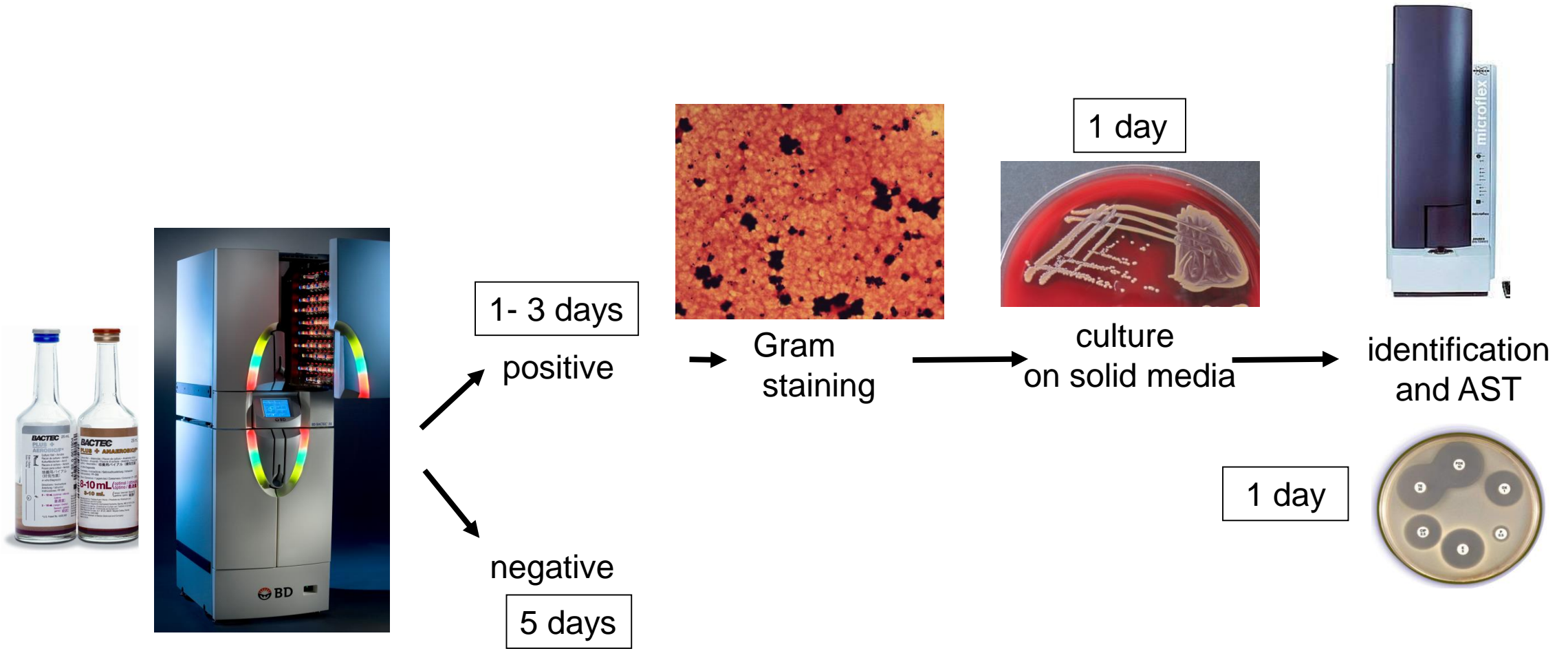


## Conclusions

- Reasons for the use of molecular approach in diagnostics:
  - speed (extreme as POCT)
  - sensitivity, specificity
  - detection of non-culturable agents
  - detection while on antibiotics
  
- In principle, two types of tests:
  - pathogen specific PCR (single agent or multiplex)
  - panbacterial PCR (metagenomics in future)

When to use which?

# Examples: diagnostics of bloodstream infections



# Examples: diagnostics of bloodstream infections

DNA diagnostics

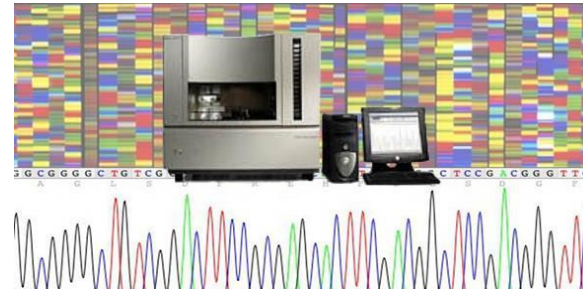
preselected multiple  
microorganisms

any microorganism



3.5 hours

*Escherichia coli*  
*Staphylococcus aureus*  
*Klebsiella pneumoniae*  
*Acinetobacter baumannii*  
*Pseudomonas aeruginosa*  
*Enterococcus faecium*



dozens of hours

*S. aureus*, *S. lugdunensis*,  
*S. epidermidis*, *S. hominis*,  
*S. haemolyticus*, *H. influenzae*,  
*S. pneumoniae*, *S. pyogenes*,  
*S. intermedius*, *S. mitis*,  
*L. monocytogenes*, *E. faecalis*,  
*E. faecium*, *E. coli*, *S. enterica*,  
*E. cloacae*, *P. stuartii*,  
*M. morgani*, *P. mirabilis*,  
*P. vulgaris*, *C. jejuni*, *C. foetus*,  
*N. meningitidis*, *B. fragilis*,  
*P. gingivalis*, *F. necrophorum*,  
*P. micros*, *F. magna*, .....

## Advantages of DNA dx

- non-culturable organisms
- detection while on antibiotics
- high sensitivity
- speed

## Disadvantages of DNA dx

- cost
- bacterial isolate not available
- missing info on AST
- not always available