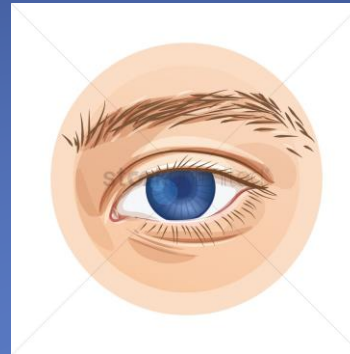
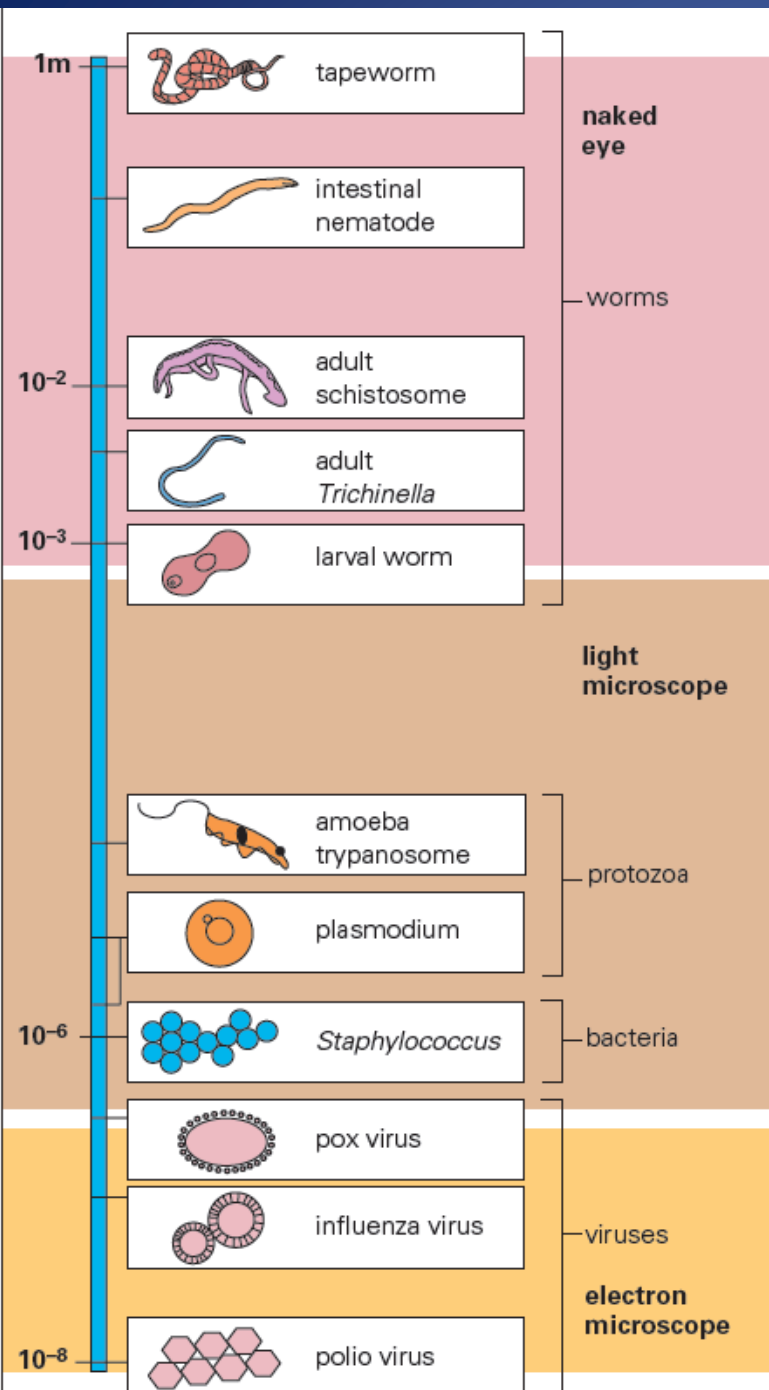


Direct diagnostics – microscopy II

Jan Tkadlec

Department of Medical Microbiology

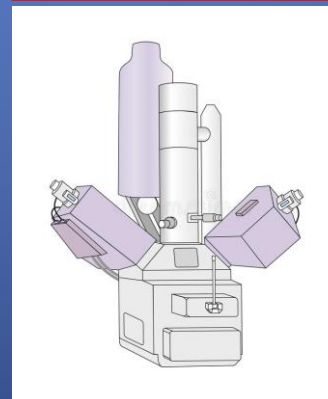
Size of human pathogens



Light microscopy

Direct observation of bacteria, yeasts, or protozoa in the sample

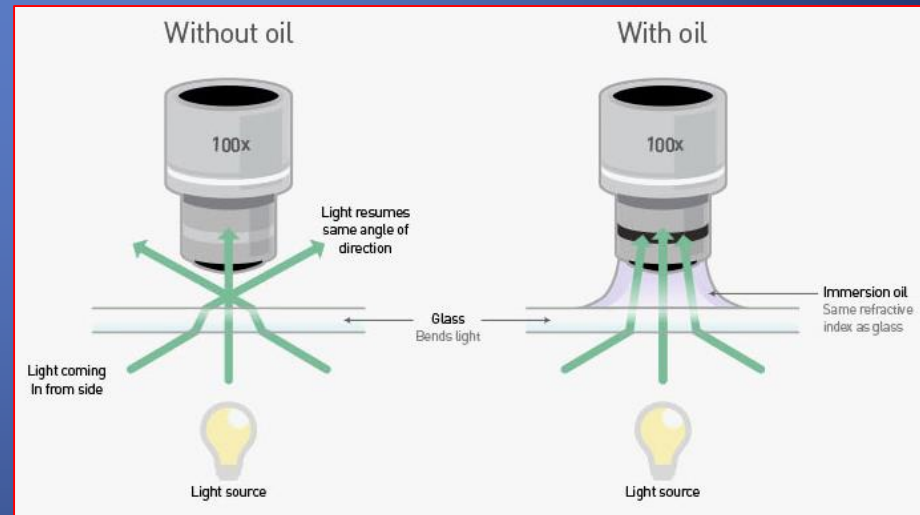
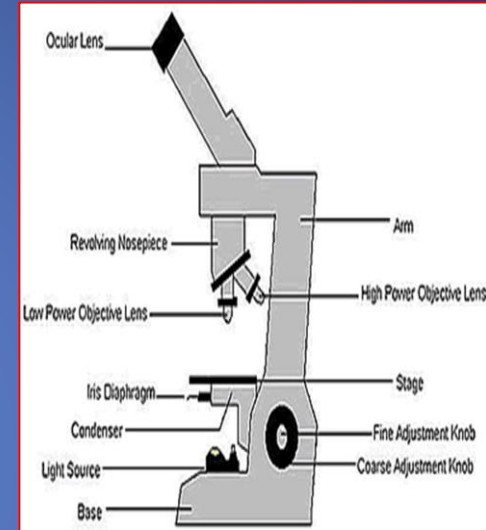
Native ~~x~~ stained sample



Electron microscope

Light microscopy

- Direct observation of microorganisms: yeasts, fungi, parasites, bacteria!
- Native (wet mount) slide – 200-400x magnification (protozoa, fungi)
- Immersion oil – up to 1000x magnific. (bacteria)



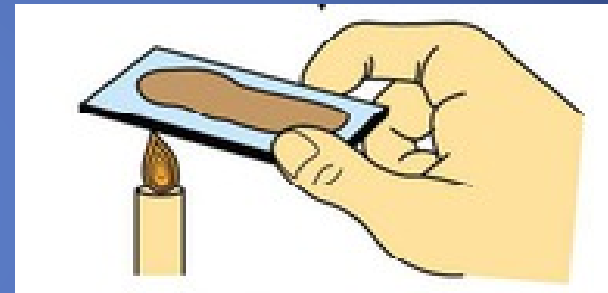
Remember

Before you start:

1. Dry



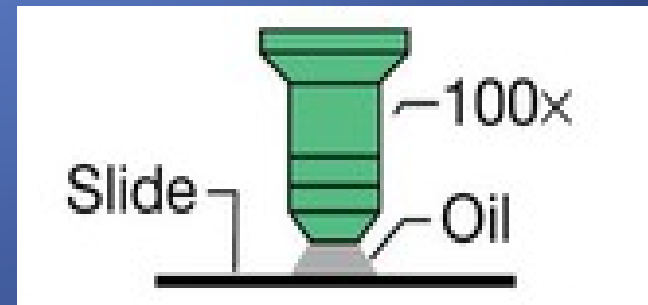
2. Heat fix



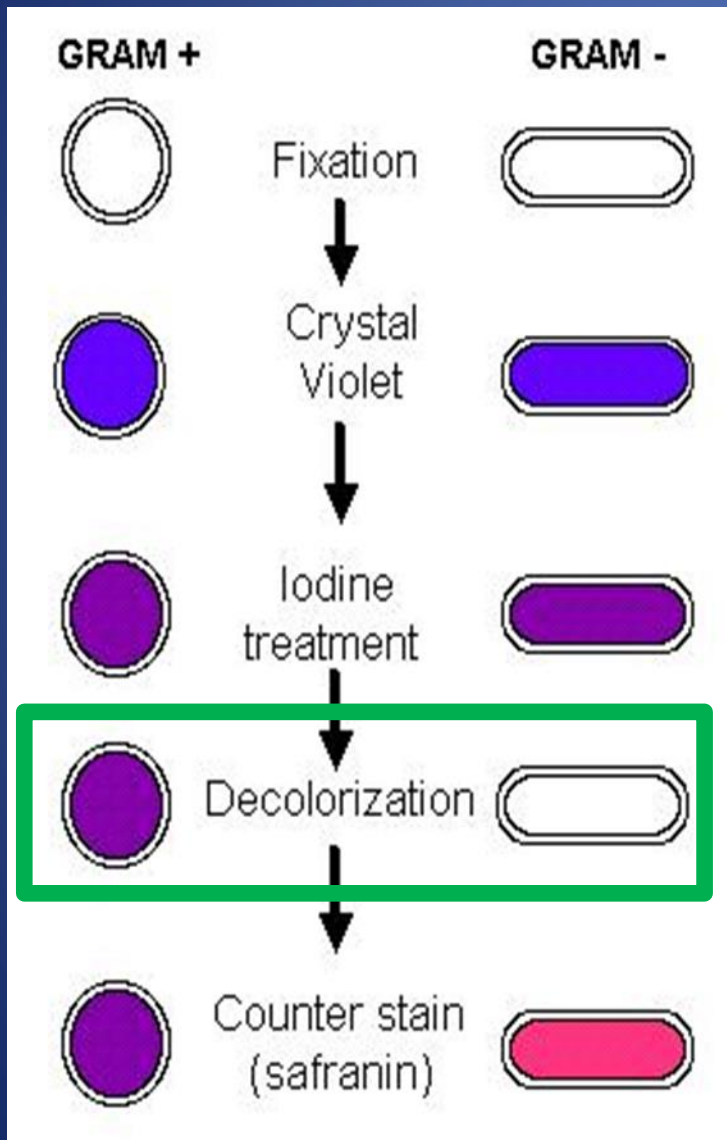
3. Stain



4. Add immersion oil

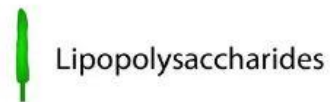
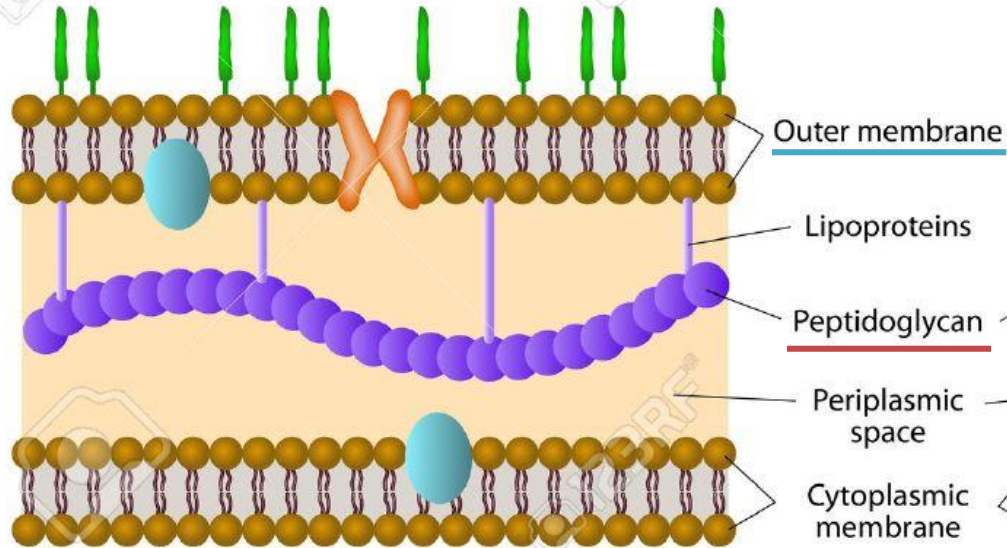


Gram stain

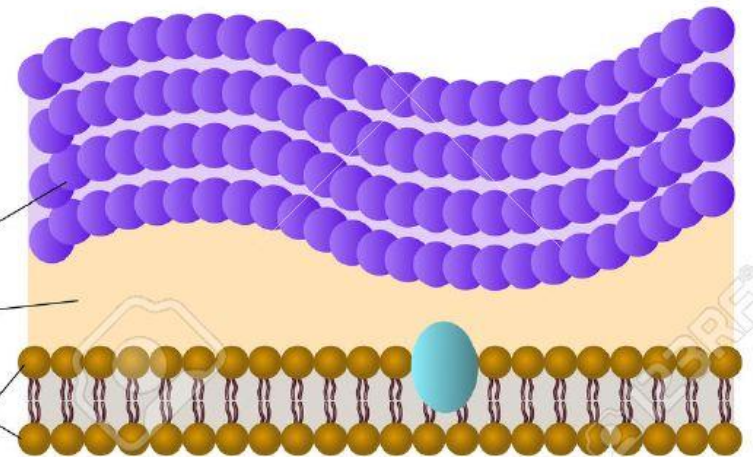


- 1. crystal violet, rinse with water
- 2. Iodine (Lugol), rinse with water
- 3. decolorization with acetone, rinse with water
- 4. counterstaining with carbolfuchsin/safranin, rinse with water
- 5. air-drying
- 6. observation with immersion oil and 100x magnif.
- G+: blue/violet, G-: pink/red

GRAM-NEGATIVE



GRAM-POSITIVE



Gram-negative:

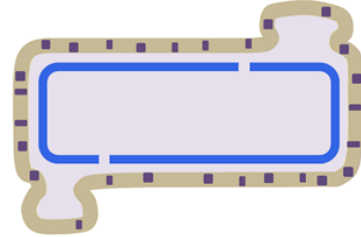
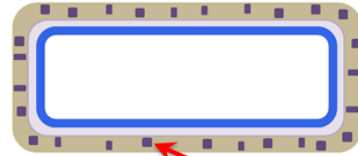
- outer membrane (not important for staining)
- Thin peptidoglycan layer!!!

Gram-positive:

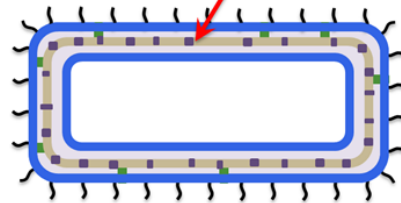
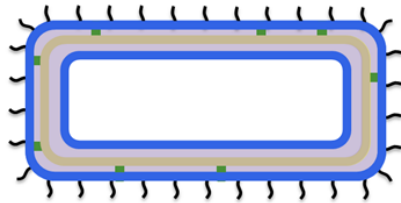
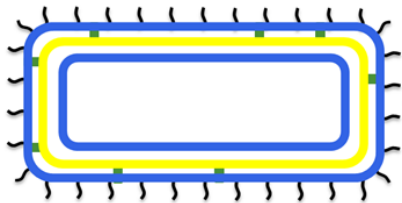
- Thick peptidoglycan layer!!!

Gram - principle

a. Gram-Positive



b. Gram-Negative



+ CV

+ Mordant

+ Alcohol wash

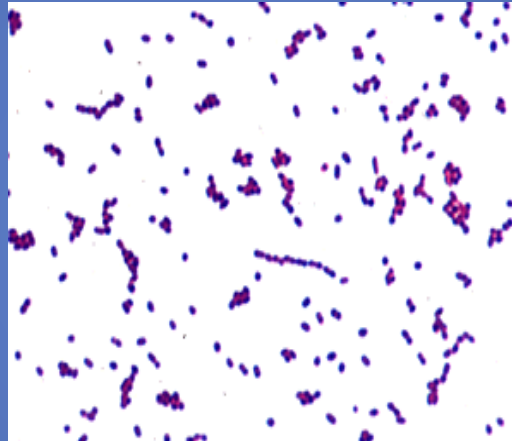
ppt.

The addition of **Crystal violet** (CV) and mordant (**Lugol=iodine**) results in the generation of a precipitate (ppt.) which is largely isolated in the cell wall. Following the destructive **alcohol wash**, the intact cell wall of the Gram+ cell retains the ppt. within the cell while the perforated cell wall of the Gram- cell allows the ppt. to be washed away.

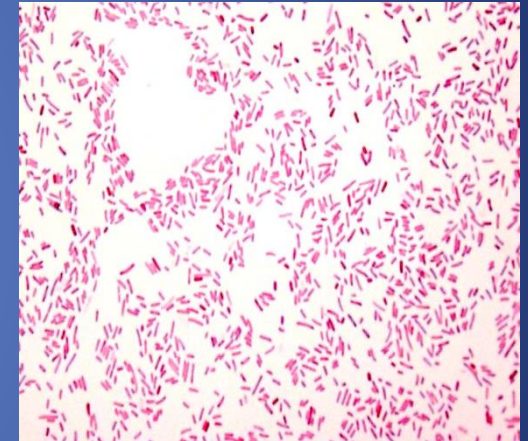
Gram - examples



G+ cocci in clusters
Staphylococcus aureus

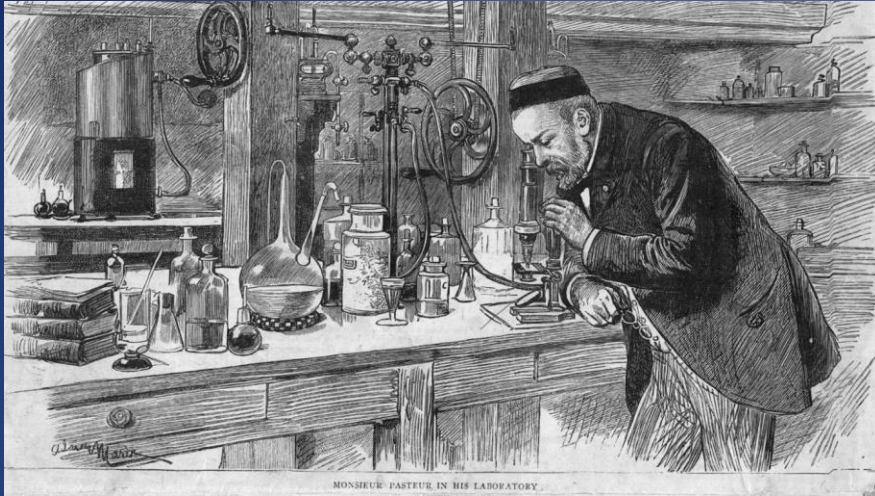


G+ cocci in chains
Streptococcus pyogenes

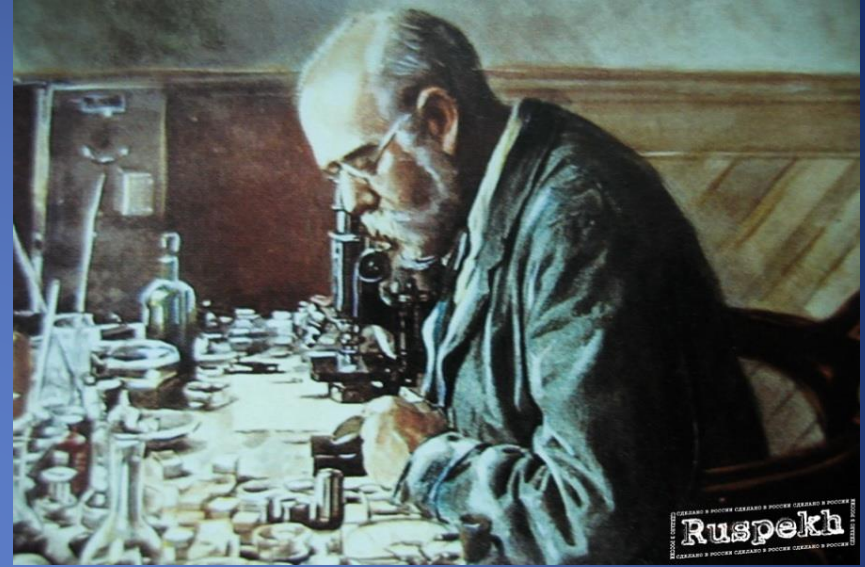


G- rods
Escherichia coli

Pasteur



Koch



Fleming



Microscope is basic tool!

Microscopy

Pros and cons

+

Speed

Low cost

Direct observation (bacteria + IS cells)

Non-cultivable organisms

-

Subjectivity

Low reproducibility

Impossible to tell exact bacterial species

Low sensitivity – negative result did not rule out infection!

Why to use microscopy:

- Identification of agents in the sample or from pure culture
 - Parasites, fungi
 - Bacteria e.g. TBC, or agents with distinctive morphology
 - Part of the biochemical identification (G+ or G-) when MALDI-TOF is not available or possible to use
- Sample validation
 - Presence (quantity) of leucocytes
 - Absence of squamous epithelia

Samples

- Clinical samples
 - liquid
 - CSF
 - Sputum and other respiratory specimen
 - Pus, discharge etc
 - Stool NOT bacteria but parasites and viruses (electron microscopy)
- Pure cultures
- Positive Blood Cultures (BC)

Blood culture diagnosis

Incubation



5-24h

Microscopy and culture on solid media



Microscopy:

- Confirmation of positivity – we see the bacteria
- Preliminary identification – guidance for a treatment

+18-24h

Identification of grown colonies



+18-24h

Antimicrobial sensitivity testing



+ 18-24h

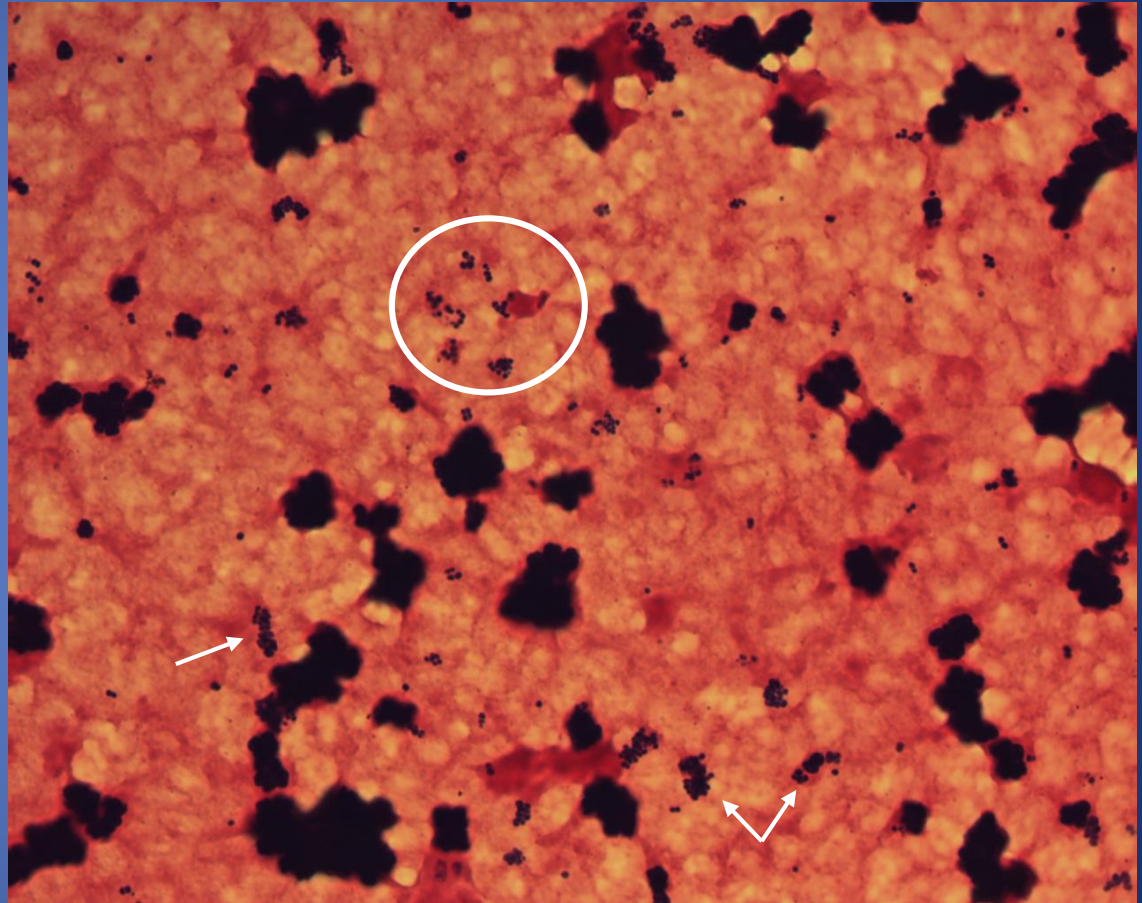
BC:
Staphylococcus
aureus

Materiál: hemokultura

Pacient: žena 84 let

Dg: endokarditida

Mikroskopie: grampozitivní koky tvořící hrozny



Autor: MUDr. Petra Kabelíková

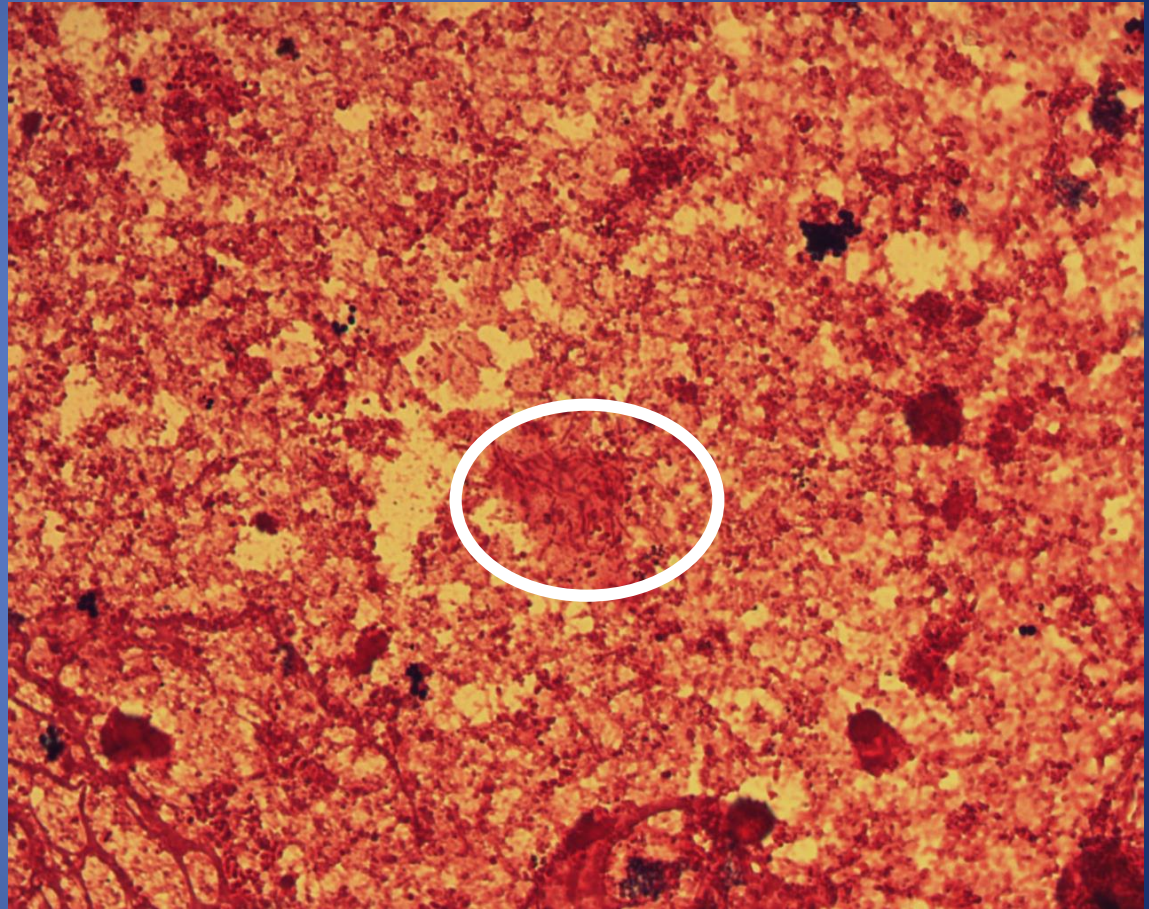
BC: Escherichia coli
Proteus mirabilis

Materiál: hemokultura

Pacient: žena 89 let

Dg: urosepse

Mikroskopie: gramnegativní tyčinky, v moči
nález *E. coli* a *P. mirabilis*



Autor: MUDr. Petra Kabelíková

Neuroinfection

CSF examination

Cytology and biochemistry: indicators of bacterial infection

turbidity

↑WBC count (PMN)

↓glucose

↑protein

Microbiology

Sample centrifugation

Microscopy

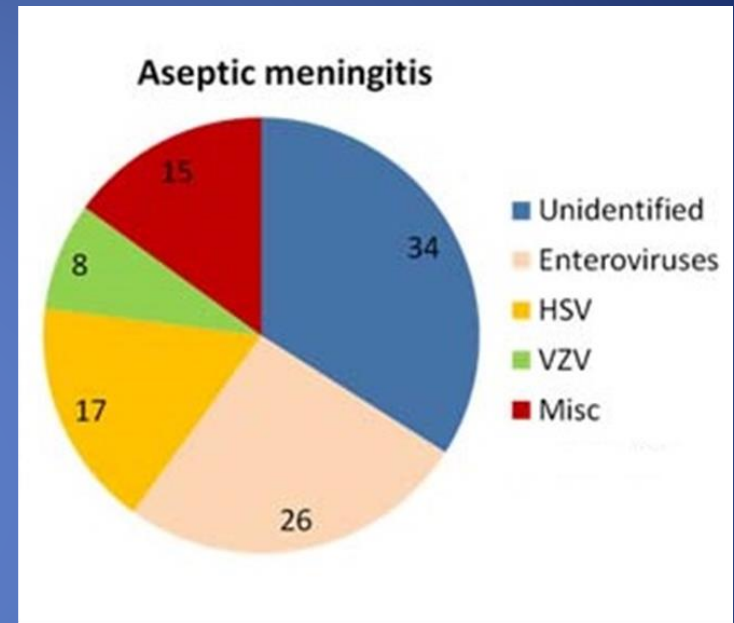
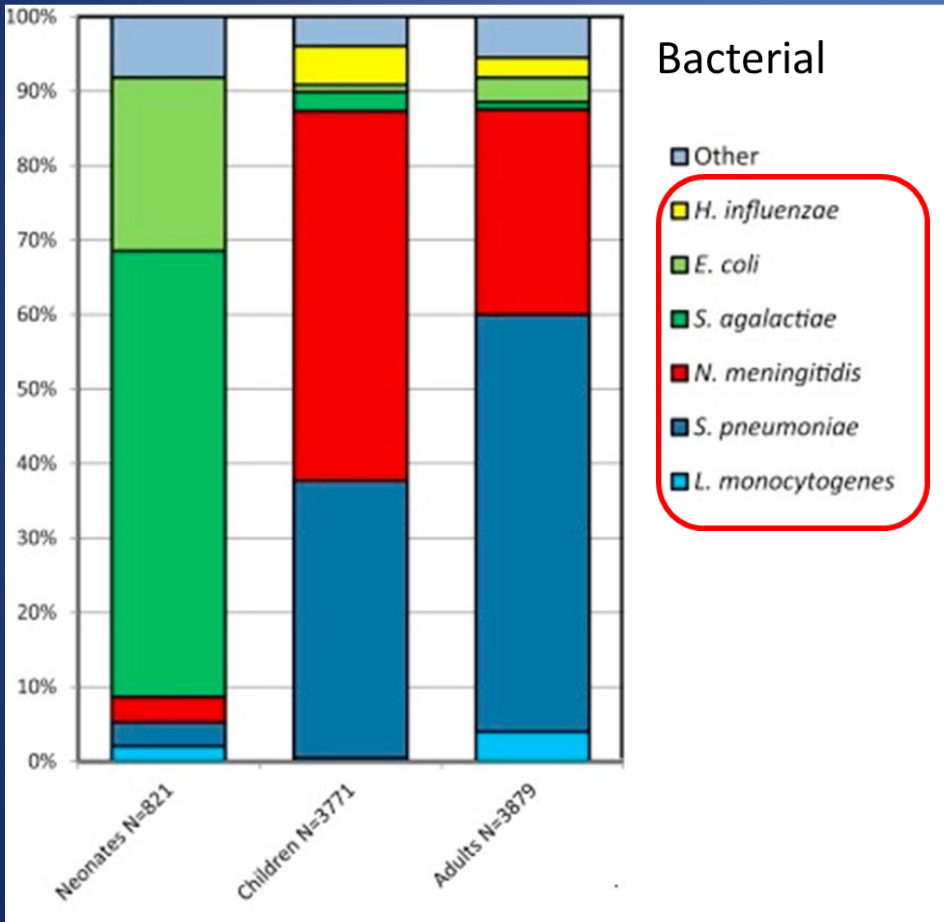
Antigen detection

PCR

Culture

CSF = cerebrospinal fluid

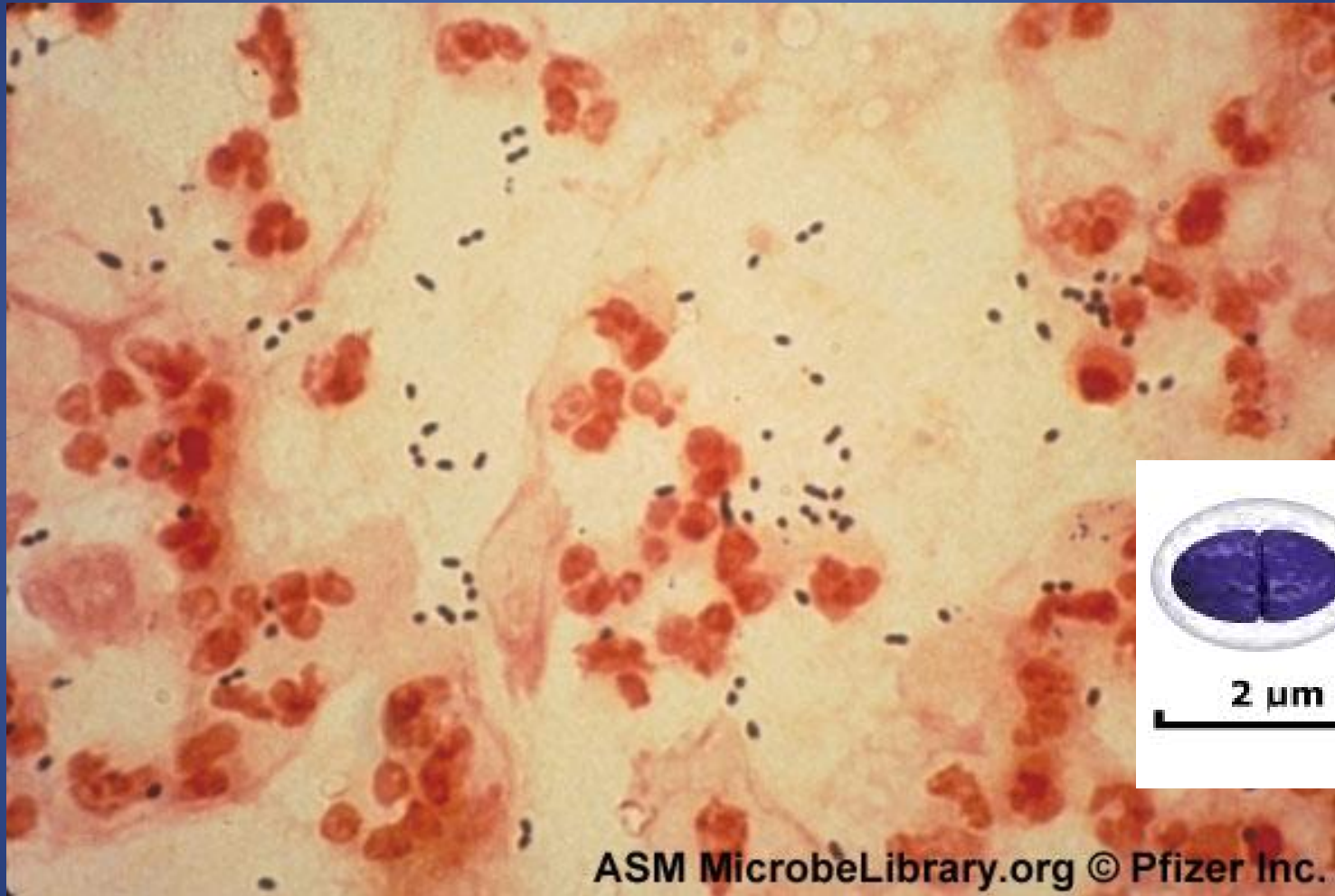
Neuroinfection



Viral agents are not visible by light microscopy

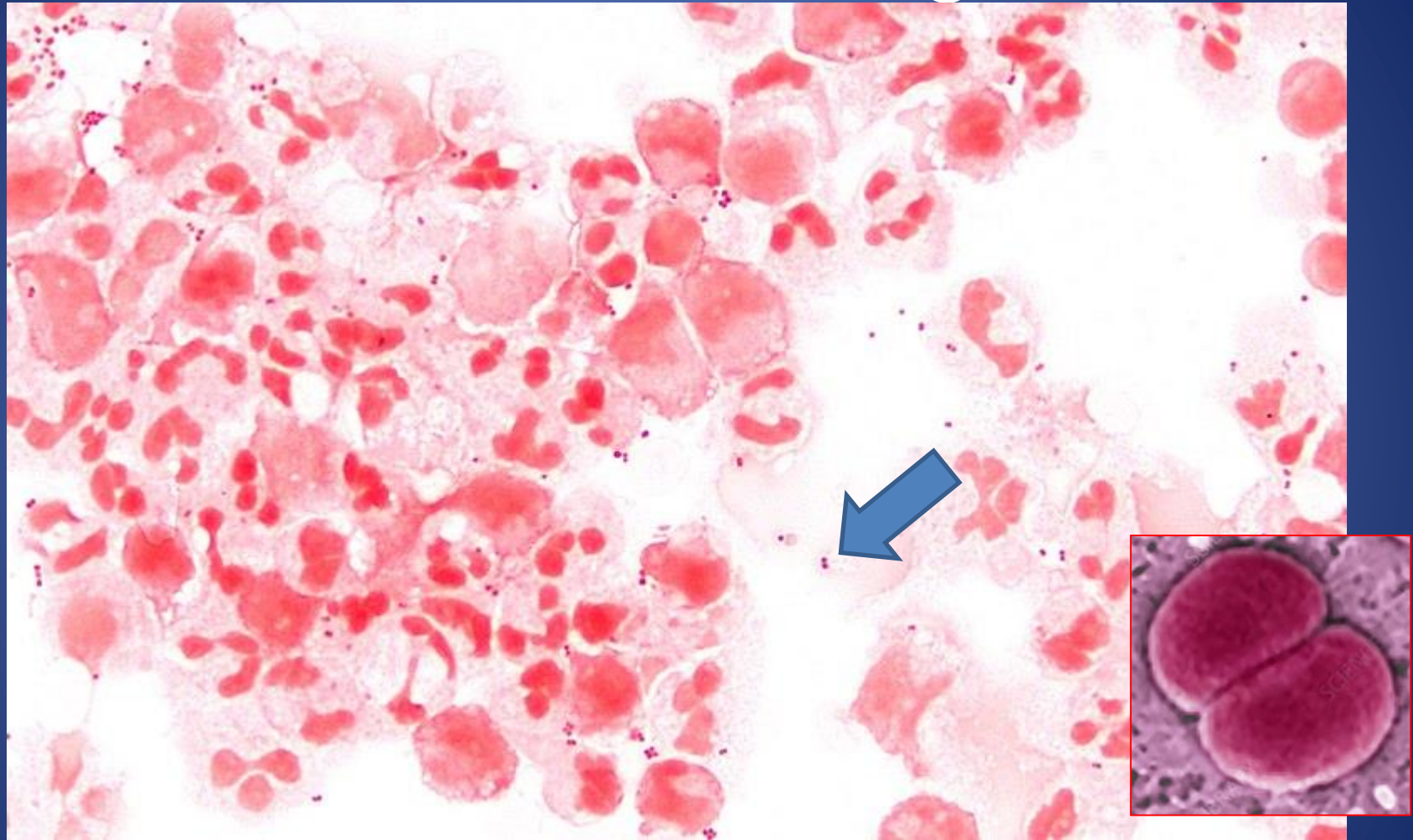
Typical morphology of main agents of septic/bacterial meningitidis

CSF: *Streptococcus pneumoniae*



Neutrophils and oval gram-positive cocci singly and in pairs (diplococci)

CSF: *Neisseria meningitidis*



gram-negative diplococci (shape of the coffee beans)

CSF: *Haemophilus influenzae*



gram-negative coccobacilli, primarily in the cytoplasm of the white cells

CSF: *Listeria monocytogenes*



Typicaly few neutrophils and few bacteria – grampositive rods (bacili)

Case report culture negative *N. meningitidis* CSF infection

- Young girl, 5 months, visiting Emergency with fever and spreading rash (purpura fulminans) – suspicion of meningococcal sepsis/meningoencephalitis
- Empirical therapy with Ceftriaxon (cephalosporin) started immediately
- Bloodculture and CSF samples taken after administration of ATB
- Microbiology :
 - BC negative, CSF culture negative (due to therapy)
 - Diplococci in blood and CSF –
N. meningitidis
 - PCR confirmed *N. meningitidis*

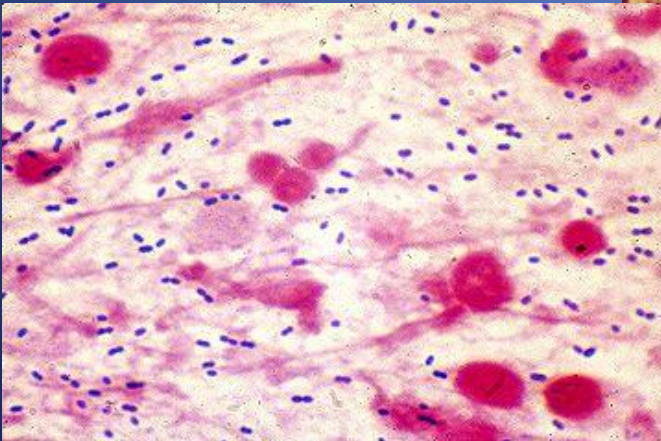
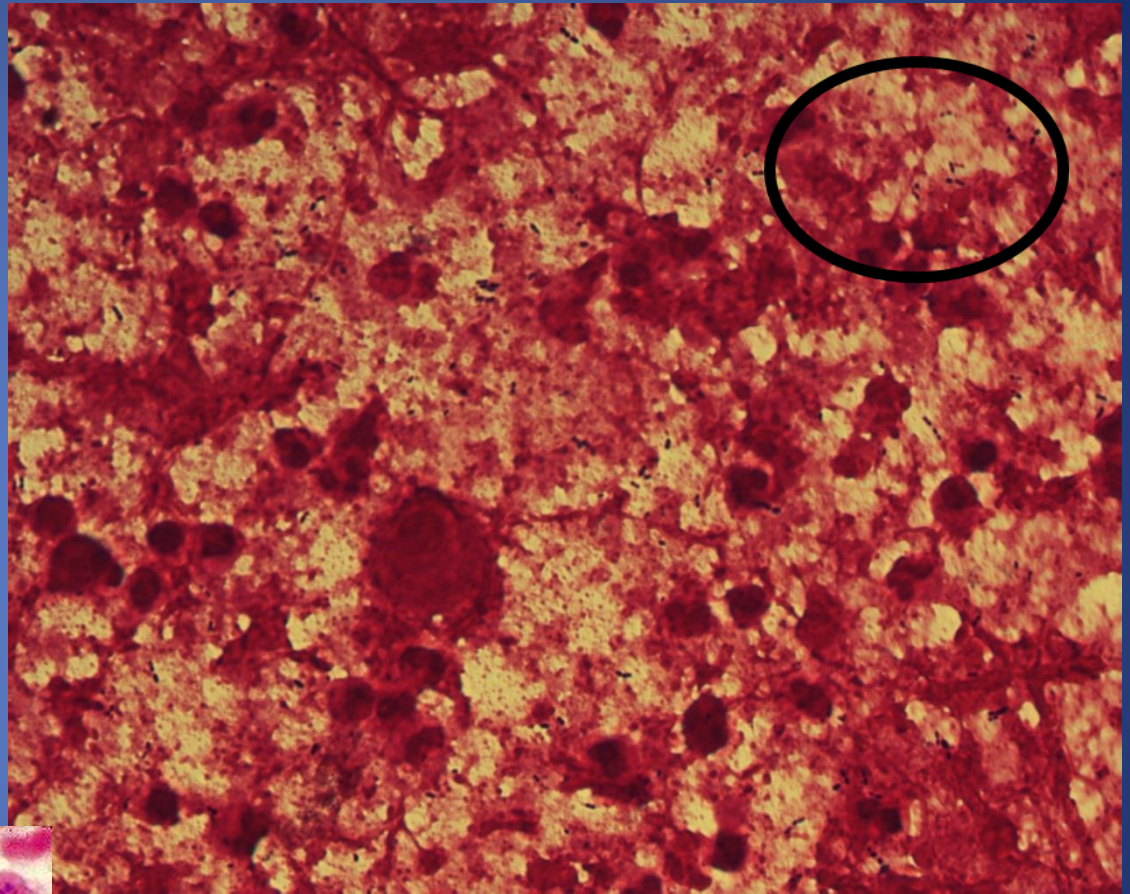


Microscopy of respiratory samples

- sputum and aspirates
- We are looking for microbes but also for neutrofiles (markers of the inflamation)
- Presence of high number of squamous epitelia (oral cavity) indicates invalide sample
- Be aware that contaminating bacteria could be present e.g. CoNS

Sputum:
Streptococcus
pneumoniae

Pneumonia



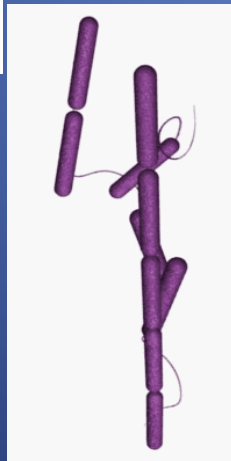
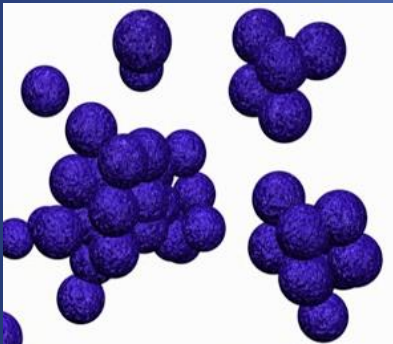
Autor: MUDr. Alena Pířová

Sputum:

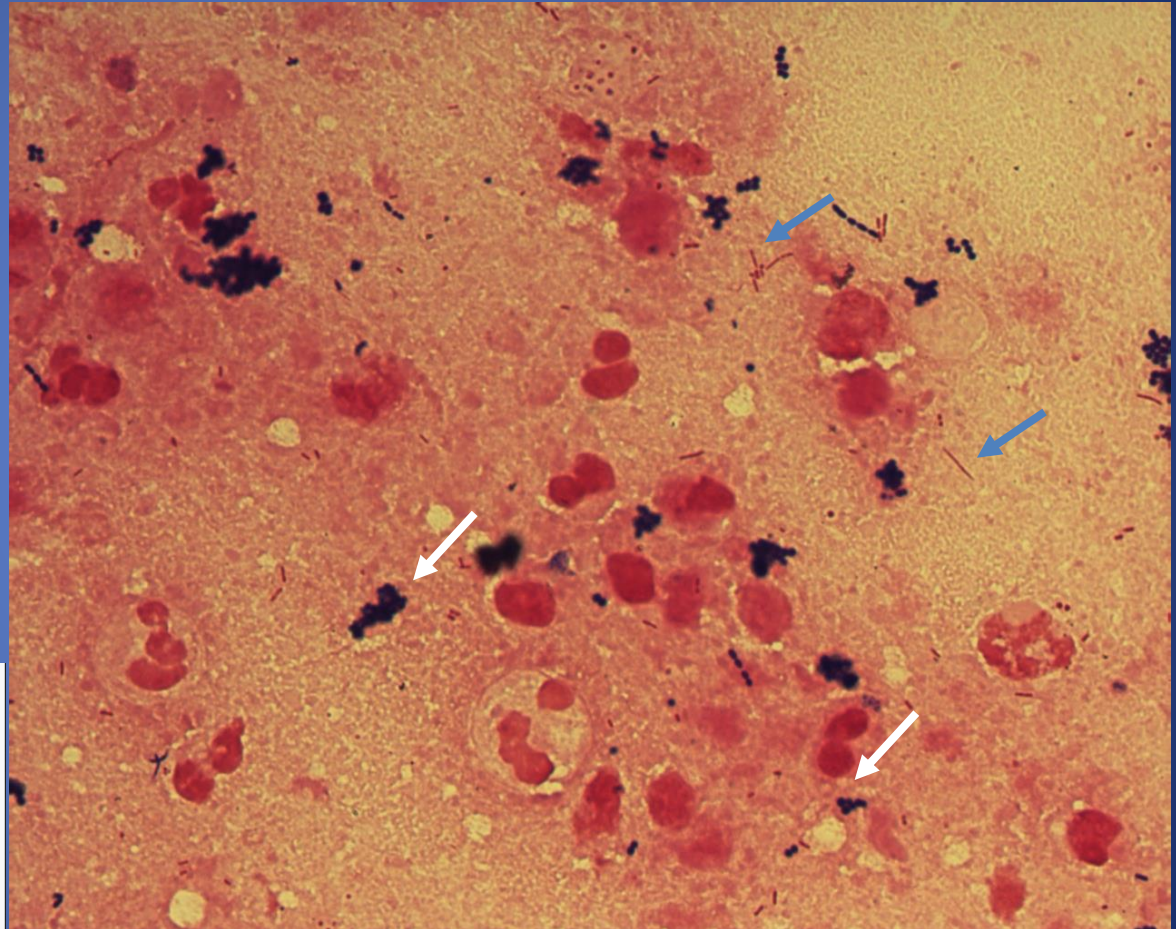
Staphylococcus aureus, *Pseudomonas aeruginosa*

Patient with cystic fibrosis

Staph. aureus



P. aeruginosa



Tuberculosis

- *Mycobacterium tuberculosis* – extremely long cultivation up to 6 weeks
- Diagnosis– microscopy (Ziehl-Neelsen) or PCR

Mycobacteria stain (Ziehl-Neelsen)

Acid-Fast Acid-Fast Negative

Cells prior to staining are transparent.



After staining with carbolfuchsin, cells are reddish-purple. Steam heat enhances the entry of carbolfuchsin into cells.



Decolorization with acid alcohol removes stain from acid-fast negative cells.



Methylene blue is used to counterstain acid-fast negative cells.



TB Stain ZN (Ziehl-Neelsen)



Capsule stain

- *indirect visualization of capsules*
- 1. a drop of India ink is placed on a side of a slide
- 2. loopful of bacteria is resuspended in ink
- 3. spread (as with blood smear)
- 4. air-dry
- 5. stain with crystal violet 1 min
- 6. rinse with water (gently, thoroughly)
 - capsules are not stained
 - Bacteria are violet
 - background is black (ink)

