

Disinfection and Sterilization Techniques, Methods of Sterility Control.

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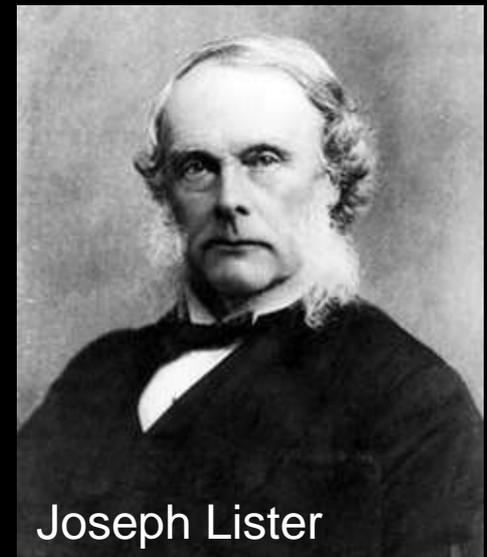
1st Faculty of Medicine



Aim:

Destruction / Removal of Harmful Microorganisms

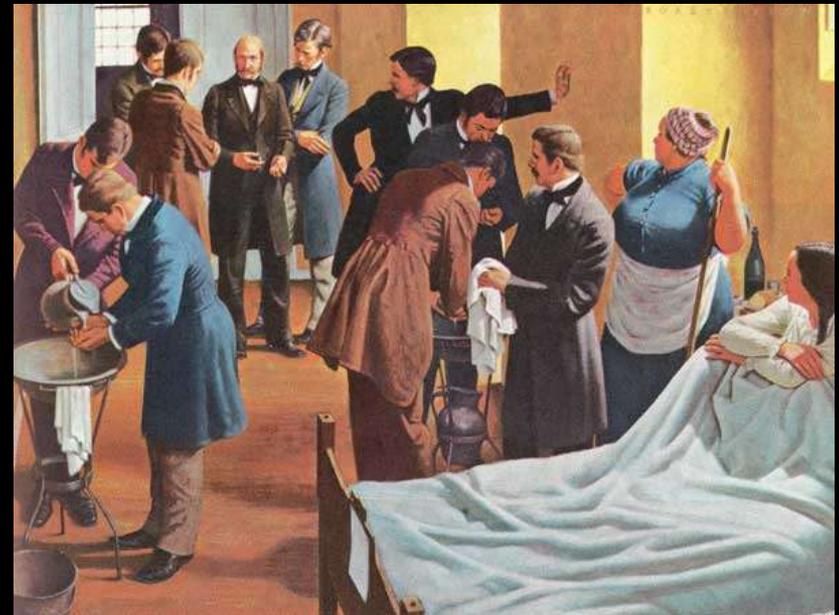
- Foundation of modern medicine
- Prevention of nosocomial infections
- Prevention of epidemics
- Lister 1865 – carbolic acid (phenol)
– antiseptic surgery



Joseph Lister

Ignaz Semmelweis (1818-1865)

- Vienna Maternity Clinics (1846)
- Incidence of death from **puerperal fever**:
1st clinic ~13% mortality (teaching of students)
2nd clinic ~ 2% mortality
- **Hand washing** with calcium hypochlorite
- Equal mortality in one year



Crises X Progress

- Major developments connected with severe health care crises
- 1970 – **Hepatitis B**: wearing of gloves, single use needles and syringes
- 1980 – **AIDS** epidemic: testing of blood and blood donors
- 2000 – prevention of **vCJD** spread (**prions**)



Guideline for Disinfection and Sterilization in Healthcare Facilities, 2008

William A. Rutala, Ph.D., M.P.H.^{1,2}, David J. Weber, M.D., M.P.H.^{1,2}, and the Healthcare
Infection Control Practices Advisory Committee (HICPAC)³

<https://www.cdc.gov/infectioncontrol/guidelines/disinfection/index.html>

Definitions

- **Cleaning** – process which removes contamination but does not destroy micro-organisms
- **Disinfection** – using an agent that destroys harmful (pathogenic) microbes, usually referred to chemicals that kill the vegetative forms but not the resistant spores of bacteria
- **Antisepsis** – disinfection on living tissue
- **Sterilisation** – process that will destroy all forms of life, including bacterial, fungi, spores, and viruses (prions)

Methods

- **Physical**

- Heat
- Filtration
- Irradiation
- Quarantine

- **Chemical**

- Choice of method depends on purpose
- Different levels of object disinfection
 - **CRITICAL**
 - **SEMICRITICAL**
 - **NONCRITICAL**
- Cleaning precedes any method

Requirements

- **CRITICAL** - objects which enter normally sterile tissue or blood should be **sterile**
(surgical instruments; cardiac catheters; implants)
- **SEMICRITICAL** - objects that touch mucous membranes or skin that is not intact require **high-level disinfection** – HLD (kills all microorganisms but high numbers of spores)
(respiratory therapy equipment; GI endoscopes)
- **NONCRITICAL** - objects that touch only intact skin require **low-level disinfection**
(bedpans; EKG leads; furniture; walls; floors)

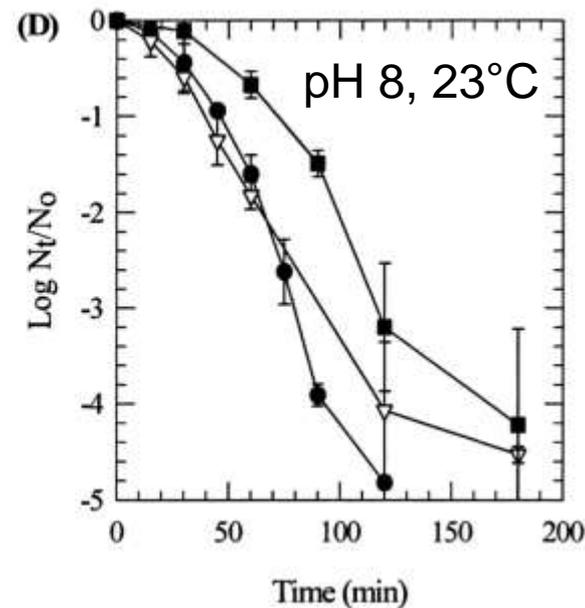
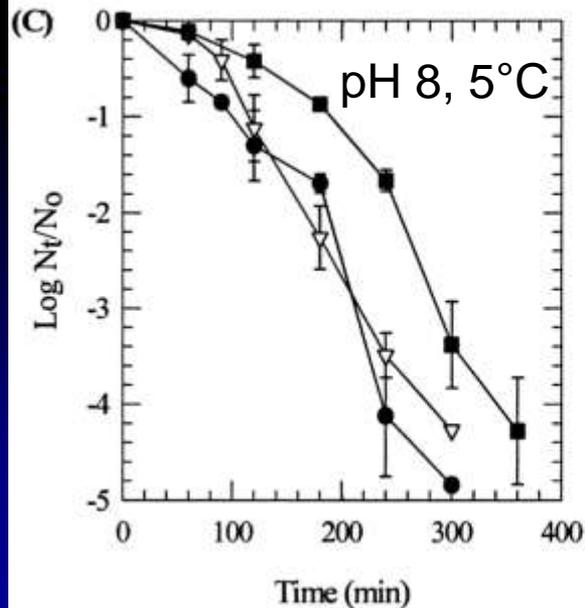
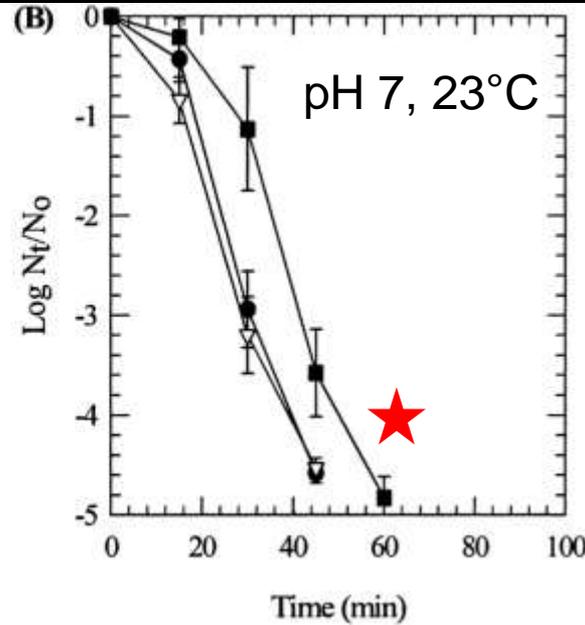
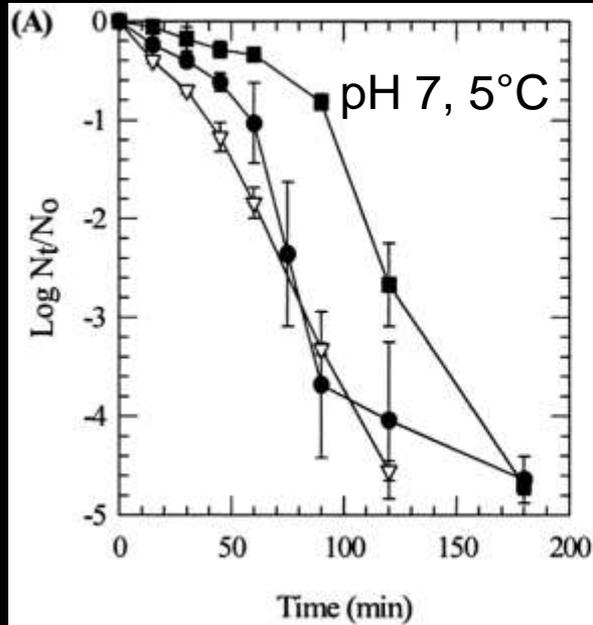
The Ideal Disinfectant

- **Broad activity** (against different pathogens)
- **Not poisonous** (or otherwise harmful)
- **Penetrating** (to pathogens)
- **Not damaging** to materials
- **Stable** and resistant to inactivation
- Easy to work with
- Otherwise not unpleasant

... difficult to fulfill

Factors influencing performance

- Strength of the disinfectant (**concentration**)
- Length of the exposure (**time**)
- **Temperature** of environment
 - rate of microbe death doubles with every 10°C rise in temp.
- Substrate effect (**contamination**)
 - level of contamination e.g. by blood or soil
- **Number of microbes** to be killed
- **Type of infectious agent**



Inactivation of *Bacillus* species spores exposed to 2.0 mg/liter free chlorine under different conditions:

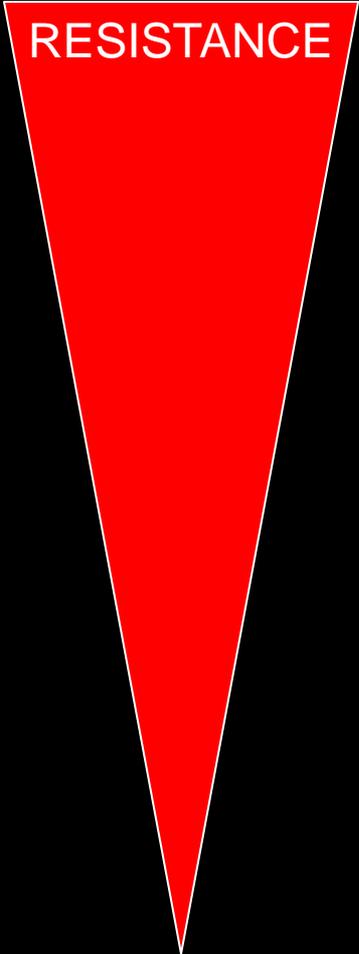
- min.
- (A) pH 7, 5°C: 150
- (B) pH 7, 23°C: 60
- (C) pH 8, 5°C: 350
- (D) pH 8, 23°C: 180

Symbols:

- B. anthracis*
- B. cereus*
- B. Thuringiensis*

Decreasing order of resistance to disinfectants / sterilants

Prions
Spores
Mycobacteria
Non-Enveloped Viruses
Fungi
Bacteria
Enveloped Viruses



RESISTANCE

Chemical disinfectants / sterilants

- **Alcohols** (ethyl alcohol, isopropyl alcohol)
- **Halogens** (iodophors and chlorine compounds)
- **Aldehydes** (formaldehyde, glutaraldehyde)
- **Ethylene oxide**
- **Peroxides** (hydrogen peroxide, peracetic acid)
- **Phenolics**
- **Detergents** (soap, quaternary ammonium comp.)
- **Heavy metals** (silver, copper)
- **Acids**
- **Hydroxides** (sodium hydroxide)

Alcohols



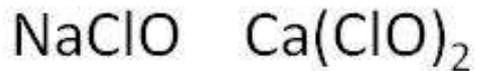
- Ethanol, Isopropanol
- Disrupt membranes, denature proteins
- Kill vegetative bacteria in 30 seconds
- Inactive against spores, fungi
- Correct dilution (~ 70%)
- Inflammable



Practical uses:

- Skin antisepsis
- Hand disinfection
- Surface disinfection





Halogens



oxidizing agents, damage membranes

Hypochlorites, chlorine

- Activity: bacteria, fungi, spores, viruses, prions
- Unstable if diluted (0.1- 2 %)
- Easily inactivated
- Corrosive for metals

Practical uses:

- Disinfection of water
- Surface disinfection (household bleach ~5 %)



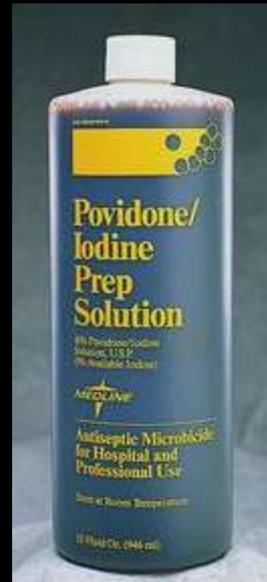
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Iodophors, iodine

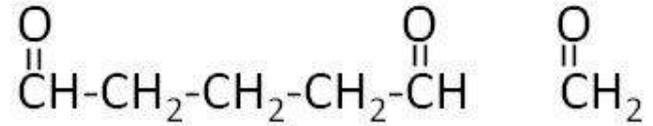
- Intermediate activity
- Good penetration
- Iodophors; + solubilizer e.g. povidone-iodine
- Easily inactivated
- Stain skin, irritant

Practical uses:

- Skin antiseptic
- Surgical scrub



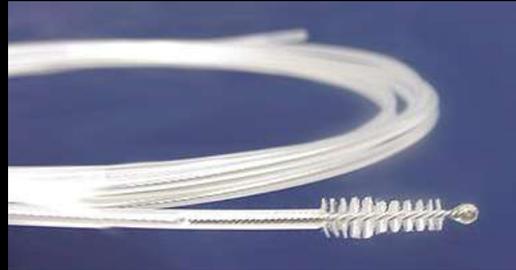
Aldehydes



- **Glutaraldehyde, formaldehyde**
- Alkylating agents
- Broad activity: bacteria, fungi, spores, viruses
- Irritant and Toxic ! Washing !!!
- Unstable – freshness and pH critical
- **ortho-phthalaldehyde** – less irritating
- Practical uses:
 - HLD or sterilization at low temperature
 - Disinfection of endoscopes
 - Fumigation of rooms



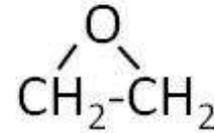
Colonoscop - HLD



Washing of endoscope after HLD



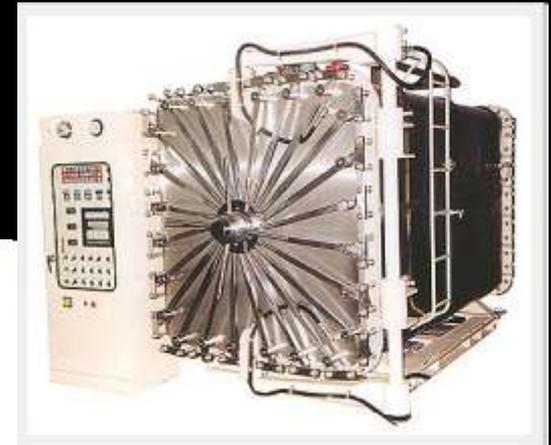
Ethylene oxide



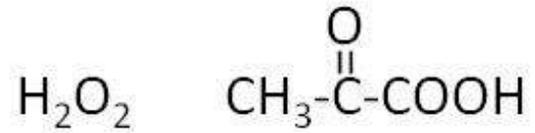
- **colorless gas**, alkylating agent
- Broad activity (not prions)
- Good penetration even in plastic
- Flammable, explosive and toxic !!
- Lengthy procedure, costly, danger

Practical uses:

- Sterilization of moisture and heat sensitive critical items

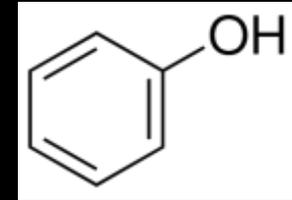


Peroxides



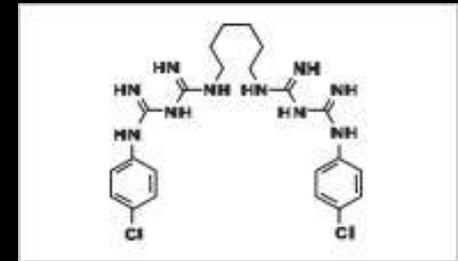
- **Hydrogen peroxide, peracetic acid**
- Oxidize organic molecules
- Broad activity: bacteria, fungi, spores, viruses
- No odor or irritation issues, nontoxic waste
- Effective in the presence of organic matter
- Can damage eye and skin, material compatibility
- Practical uses:
 - HLD or sterilization at low temperature
 - Disinfection of endoscopes
 - Antiseptics (3% H_2O_2), contact lenses

Phenolics



- **Phenol, phenol derivatives**
- Precipitate proteins, damage membranes
- Intermediate activity, not sporicidal
- Use: disinfectants on soiled surfaces

Detergents



Chlorhexidine

- **Soap, quaternary ammonium compounds**
- Damage membranes, enzymes
- Low to intermediate activity
- Use: disinfectants for noncritical objects, skin

Heavy metals

- **Silver, copper**
- Oligodynamic effect, inactivate enzymes
- Use: AgNO_3 prophylaxis of newborn conjunctivitis, silver coating of catheters, Cu fungicides

Acids and Hydroxides

- **Sodium hydroxide**
- Strong disinfectant, alkalic hydrolysis
- Not inhibited by organic contamination
- Use: destruction of prions (1M NaOH, 60 min.)

General disinfection rules

- Choose appropriate disinfectant, rotate (!)
- Use **correct concentration** (follow instructions)
- Do **not use old solutions** (> 24 h)
- Do not top up old solutions
- **Correct length, temperature and pH** of treatment (instructions)
- Control sterility of disinfectant itself (e.g. *Pseudomonas* sp. in QAS)

Physical methods

- Heat
 - dry
 - moist
- Filtration
 - liquids
 - gases
- Irradiation
 - UV
 - gamma
- Quarantine / Cover

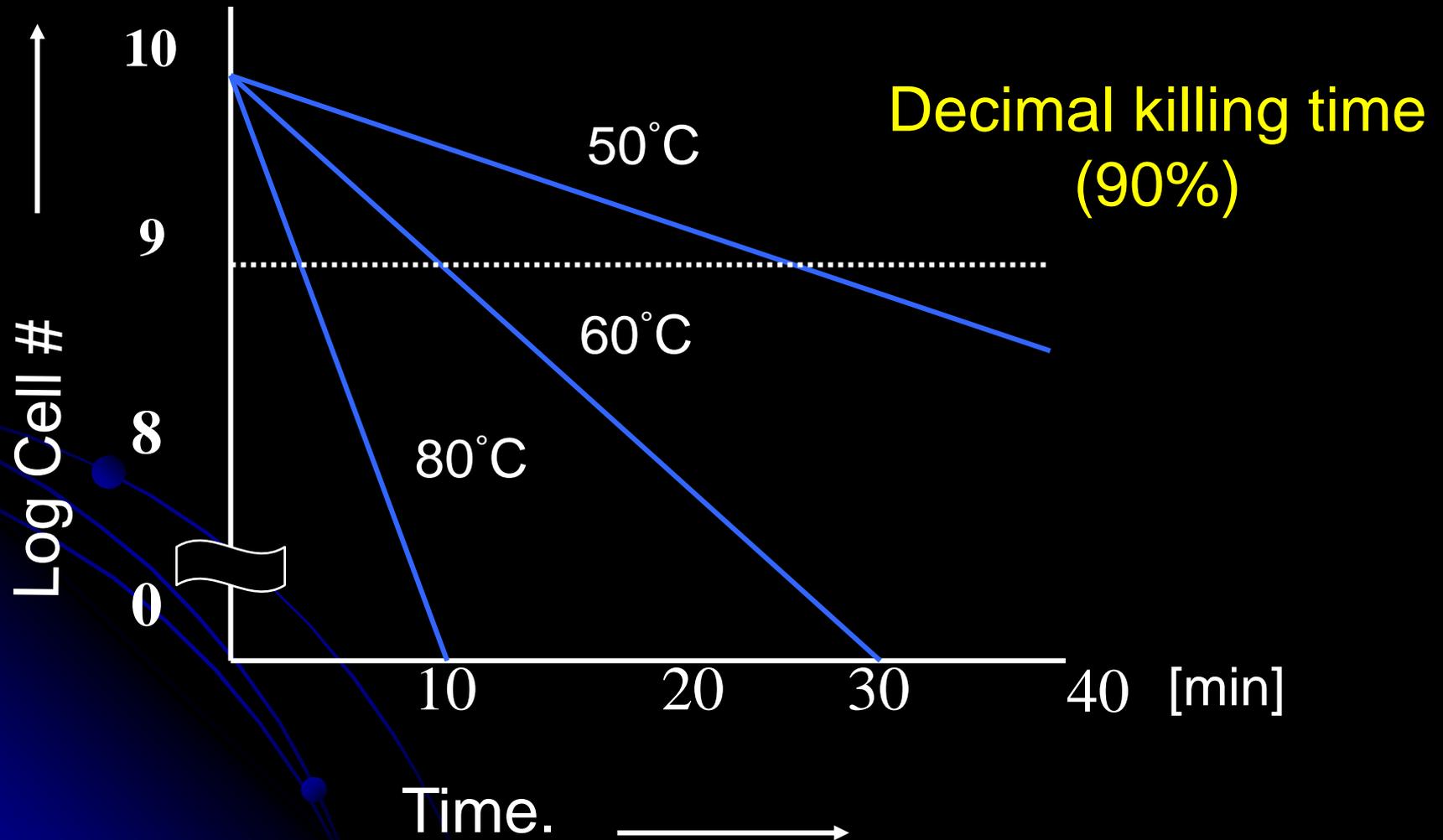


Heat

- Nontoxic, quick, cheap
- Only for heat resistant items
- Pasteurization – kills vegetative bacteria (e.g. 72°C, 20 sec.)
- Boiling
- Flaming (microbiology loops)
- Incineration – the ultimate sterilization (medical and biohazardous waste)



Logarithmic lethality curve



Dry heat sterilization

- **Hot air sterilizer**
- Oxidation of cellular components
- For moist sensitive materials (e.g. oils, powders, sharp instruments)
- Non corrosive
- **150-170°C (150 - 60 min)**



Steam sterilization

- **Autoclave**

- Denature enzymes and proteins
- Saturated steam applied under pressure
- Direct contact with items
- Nontoxic, rapid, cheap and dependable
- Exclusion of air from the chamber

Gravity displacement autoclave (121°C, 20 min.)

High pre-vacuum autoclave (121°C, 4 min.)

Autoclaves

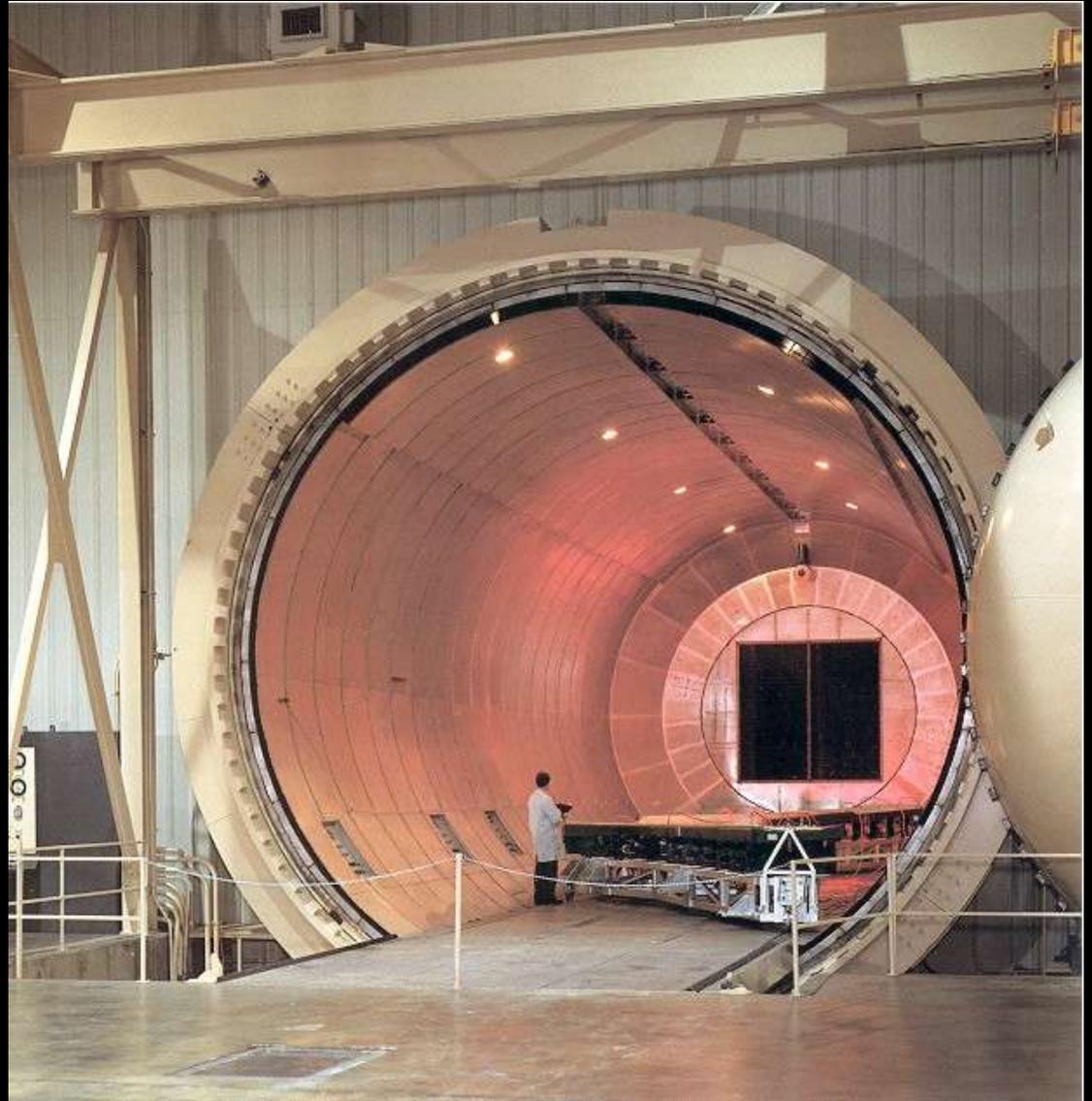
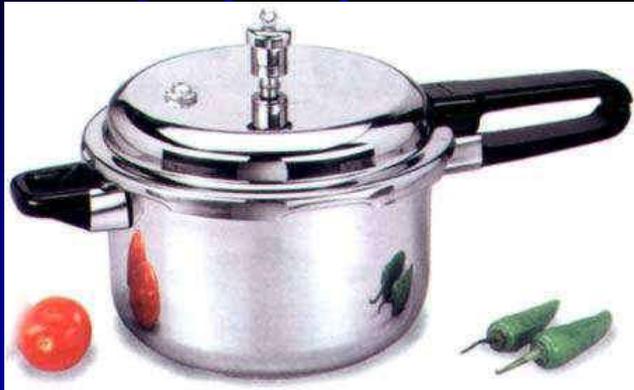


Autoclave large



Autoclave for space ship

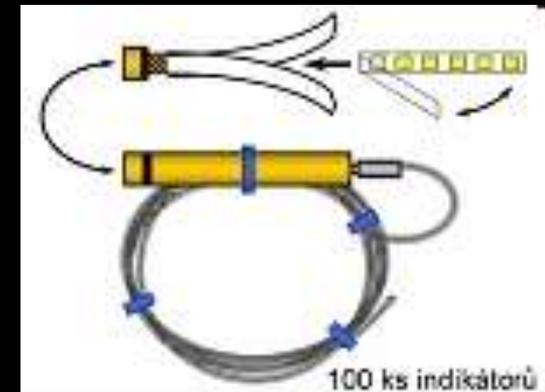
Pressure cooker



Autoclaving

- **Controlled cycle** (temperature, pressure, time)
- **Typical settings** (gravity displacement):

121°C	200 kPa	20 min
134°C	300 kPa	10 min (prions 60 min)
- **Monitoring of effectiveness**
 - temperature & pressure charts
 - chemical indicators (e.g. Bowie-Dick test)
 - biological indicators (spore tests, *B. stearothermophilus*)
- **Documentation** (store 15 years)



Filtration – liquids & gases

- **Ultrafiltration** of liquids – pore size e.g. 0.2 μm
- Labile fluids



- **Gases (Air)** – HEPA or ULPA filters
- Biosafety cabinets
- Infectious disease labs (e.g. BSL-3)
- Infectious disease clinics



Irradiation

- **UV irradiation** (220 – 330 nm)
- Causes DNA damage (254 nm), germicidal
- Disinfection of surfaces, water

- **Gamma irradiation** (^{60}Co , 25 kGy - high energy)
- Causes DNA damage
- Only in specialised centres
- Used on disposable plastics in sealed packs



Low temperature plasma

- **Hydrogen peroxide gas plasma**
- Generated using radio frequency or microwave energy – reactive particles, free radicals
- Cycle ~ 60 min. at 45°C
- Broad activity including spores (and prions??)
- Nontoxic
- For thermo-labile materials (compatible with >95 % of medical devices)



Sterilization summary

CHEMICAL METHODS

- Glutaraldehyde (> 2.0%)
- Hydrogen peroxide (7.5%)
- Peracetic acid (0.2%)
- Ethylene oxide

PHYSICAL METHODS

- Heat
- Filtration
- Irradiation

NEW METHODS

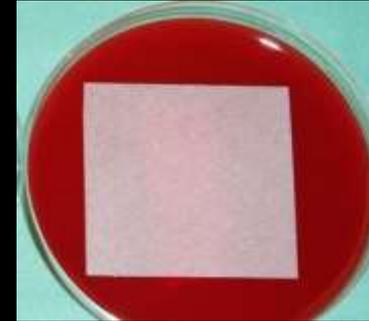
- Low temperature plasma

Pre-sterilization: cleaning (washing), drying, wrapping

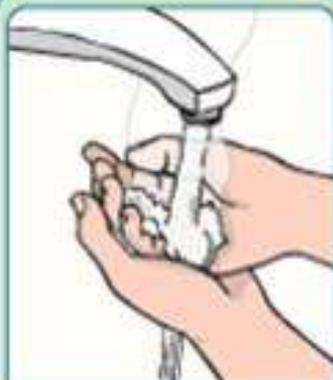
Environmental microbiological control

- **Surfaces** - swabbing, printing
- **Air** - sedimentation
- **Liquids** - direct cultivation
- **Materials** - rinsing, immersion

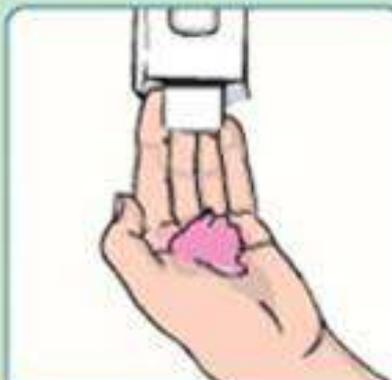
Cultivation detection, molecular biology, immunomethods..



FIGHT GERMS BY WASHING YOUR HANDS!



1 Wet your hands



2 Soap



3 Lather and scrub - 20 sec



4 Rinse - 10 sec



5 Dry your hands



6 Turn off tap

DONT FORGET TO WASH:

- between your fingers
- under your nails
- the tops of your hands

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